## HIV-Associated Cardiovascular Disease Pathogenesis: An Emerging Understanding Through Imaging and Immunology

Jo[n](https://orcid.org/0000-0002-5128-8388)athan A. Hudson<sup>o</sup>[,](https://orcid.org/0000-0001-9119-3167) Rashida A. Ferrand<s[u](https://orcid.org/0000-0002-8908-4735)p>o</sup>, S[a](https://orcid.org/0000-0003-3926-4225)muel N. Gitau<sup>o</sup>, Marianne Wanjiru Mureithi<sup>o</sup>, Pasquale Maffia<sup>o</sup>, Shirjel R. Ala[m](https://orcid.org/0000-0002-2099-612X)<sup>D</sup>,\* Anoop S.V. S[h](https://orcid.org/0000-0002-2825-3419)ah<sup>D\*</sup>

**ABSTRACT:** Cardiac abnormalities were identified early in the epidemic of AIDS, predating the isolation and characterization of the etiologic agent, HIV. Several decades later, the causation and pathogenesis of cardiovascular disease (CVD) linked to HIV infection continue to be the focus of intense speculation. Before the widespread use of antiretroviral therapy, HIVassociated CVD was primarily characterized by HIV-associated cardiomyopathy linked to profound immunodeficiency. With increasing antiretroviral therapy use, viral load suppression, and establishment of immune competency, the effects of HIV on the cardiovascular system are more subtle. Yet, people living with HIV still face an increased incidence of cardiovascular pathology. Advances in cardiac imaging modalities and immunology have deepened our understanding of the pathogenesis of HIV-associated CVD. This review provides an overview of the pathogenesis of HIV-associated CVD integrating data from imaging and immunologic studies with particular relevance to the HIV population originating from high-endemic regions, such as sub-Saharan Africa. The review highlights key evidence gaps in the field and suggests future directions for research to better understand the complex HIV-CVD interactions.

**Key Words:** cardiomyopathies ■ coronary artery disease ■ heart failure ■ inflammation ■ ventricular dysfunction, left

The scale-up of antiretroviral therapy (ART) has<br>transformed HIV infection from an invariably fatal<br>condition to a chronic manageable carriage. Life<br>expectancy among people living with HIV (PLWH) he scale-up of antiretroviral therapy (ART) has transformed HIV infection from an invariably fatal condition to a chronic manageable carriage. Life has increased, with non-AIDS–related death rates now 4-fold higher compared with AIDS-related rates and cardiovascular disease (CVD) accounting for a large proportion of this mortality. The burden of HIVassociated CVD globally is not distributed equally with sub-Saharan Africa (SSA) suffering the greatest impact (Figure 1).1–5

The current paradigm is that atherosclerotic coronary artery disease (CAD) driven by vascular inflammation is the primary pathology of HIV-associated CVD resulting in ischemic injury and infarction of the myocardium. This paradigm, based on studies performed predominantly in North American and European populations, has strongly influenced the contemporary clinical approach and informed guidelines that focus on addressing traditional atherosclerotic risk factors in HIV.<sup>6-10</sup> These studies have also guided recent large-scale clinical trials, showing improved outcomes across PLWH receiving lipidlowering therapy.<sup>11</sup>

However, questions arise about the applicability of the current paradigm to populations with HIV in SSA, which harbors two-thirds of the 38 million PLWH. We propose that a nonischemic inflammatory cardiomyopathic process may be a more important mechanism, particularly in populations with a low prevalence of CVD risk factors.

This review will explore the evidence for the role of nonischemic inflammatory cardiomyopathy as an important mechanism of HIV-associated CVD and, in particular, its relevance to the SSA population. It will also critically

Correspondence to: Anoop S.V. Shah, PhD, Department of Non-Communicable Disease Epidemiology, London School of Hygiene and Tropical Medicine, Room 249, Keppel St, WC1E 7HT London, United Kingdom. Email anoop.shah@lshtm.ac.uk

\*S.R. Alam and A.S.V. Shah contributed equally.

For Sources of Funding and Disclosures, see page 1560.

<sup>© 2024</sup> American Heart Association, Inc.

*Circulation Research* is available at www.ahajournals.org/journal/res

## Nonstandard Abbreviations and Acronyms



review the current understanding that HIV-associated CVD is predominantly caused by atherosclerotic coronary artery disease.

## EPIDEMIOLOGY OF CLINICAL HIV– ASSOCIATED CVD

In the pre-ART era, cardiac presentations of HIV infection were dominated by cardiomyopathy, which carried a poor prognosis.12 Individuals with HIV-associated cardiomyopathy (HIVAC) confirmed on endomyocardial biopsy were at almost 6 times the risk of death compared with death rates among individuals with other forms of cardiomyopathy.13 In the era of highly effective ART, the focus of HIV-associated CVD has shifted from a disease of the myocardium toward atherosclerotic coronary artery

disease,<sup>6</sup> predominantly driven by data from PLWH in high-income countries (Table 1).<sup>9,14</sup>

## Heart Failure

Although HIV infection is an independent risk factor for the development of heart failure, the phenotype has changed over time.16 HIVAC, characterized by increased left ventricular dilatation and reduced function, was the prevalent phenotype before the implementation of ART.17 Reductions in the prevalence of HIVAC in high-income countries were documented following widespread ART use.18 A meta-analysis encompassing 26 studies indicated a decrease in left ventricular systolic dysfunction prevalence among PLWH from 20% in the pre-ART era (before 1996) to 10% in studies conducted post-2004.19 The heart failure phenotype appears to have shifted to subclinical left ventricular systolic dysfunction and diastolic dysfunction.<sup>19</sup> Four of the 26 studies analyzed were from SSA countries.<sup>20-23</sup> Despite these changes, compared with individuals without HIV infection, PLWH have a 1.8-fold increased risk of developing heart failure.16 This risk is further exacerbated by poor HIV control even after adjusting for traditional cardiovascular risk factors.24

In SSA, as in high-income countries, HIVAC was documented in up to one-third of PLWH before the availability of ART.<sup>25</sup> Echocardiographic studies in ART-naive populations in Ghana and Rwanda reported HIVAC prevalence of 34%<sup>20</sup> and 17%,<sup>26</sup> respectively. The South African Heart of Soweto study showed that 10% of all hospitalized patients with an incident cardiac pathology were HIV positive (n=518/5328). The mean age of this cohort was 40 years, and 54% of them were receiving ART. HIVAC was the most common etiology in these patients, equating to 38% (n=196/518) of cases followed by pericardial pathology at 13% (n=65/518). Noticeably, ischemic pathology only accounted for 3%  $(n=14/518)$  of cases.<sup>27</sup> Despite these findings, the true burden of HIV-associated heart failure in the region is still uncertain as ART coverage has improved significantly in recent years.<sup>28</sup>

## Ischemic Heart Disease

Several cohort studies, alongside pooled analysis, have shown that PLWH face a significantly elevated risk of acute myocardial infarction (MI) compared with people without HIV (PWOH), even after adjusting for traditional cardiovascular risk factors.4,14,29,30 All of this evidence, based on longitudinal studies, originates from studies in North America and Europe.<sup>14</sup> This elevated risk appears to persist among individuals who consistently achieve HIV viral load levels of <500 copies/mL over time.<sup>29</sup> The body of evidence pointing toward an excess risk of MI in PLWH has culminated in the recent randomized trial to



**Figure 1. The global burden of HIV-associated cardiovascular disease.**

Cartograms showing HIV attributable disability-adjusted life-years per 100 000 people for HIV-associated cardiovascular disease.14 Each color represents a septile.

reduce vascular events in HIV trial (REPRIEVE) investigating the use of statins to improve cardiovascular outcomes in PLWH.11 Pitavastatin was shown to reduce the risk of major adverse cardiac events by 35% in PLWH at low-to-moderate risk of CVD over a median follow-up of 5 years. The REPRIEVE population consisted of 70% men with 50% of the overall population being smokers: a demographic different from the HIV population in SSA (Table 2).35 Only 15% of the REPRIEVE population was recruited from SSA, which calls into question its generalizability to the global HIV population.<sup>11</sup>

The relationship between HIV and coronary artery disease in SSA remains unknown. To our knowledge, there are no cohort studies from SSA that have investigated the association between HIV and ischemic heart disease.36 The lack of HIV-specific research in this area is compounded by the limited data in the region on the burden of coronary artery disease in the general population.37,38 Among the limited studies that have explored the relationship between HIV and coronary artery disease in SSA, the Heart of Soweto Study was a South African registry study that captured all new cases of heart disease between 2006 and 2008 presenting to a tertiary

cardiology center in Johannesburg. A diagnosis of coronary artery disease was made in 10.9% (581/5328) of the cohort, and only 2.4% of these cases (14/581) were PLWH. These results suggest a comparably low burden of HIV-associated atherosclerotic disease in this population.27,39 A recent analysis of electrocardiograms (ECGs) from a rural Tanzanian cohort indicated that PLWH were five times more likely to exhibit ischemic changes on their ECGs than to age- and sex-matched PWOH.<sup>40</sup> Although not specific, these contrasting findings highlight the importance of future studies in SSA HIV populations to determine the risks of coronary artery disease associated with HIV infection.

## MECHANISMS OF NONISCHEMIC HEART FAILURE IN HIV

The epidemiological evidence for the association between heart failure and HIV suggests a potential role for a nonischemic inflammatory cardiomyopathy that causes myocardial injury. Here, we will review the evidence derived from immunologic and advanced imaging studies that shape our understanding of the pathophysiological

**Table 1. Epidemiological Evidence for Cardiovascular Complications in PLWH**

Outcome	Study design	No. of studies	<b>HR</b>	No. of studies from SSA	Source	Comment
MI	Systematic review and meta-analysis	5	$1.79(1.54 - 2.08)$	$\Omega$	Shah et al <sup>14</sup>	HR for incident MI in PLWH compared with PWOH
Heart failure	Systematic review and meta-analysis	5	$1.48(1.31 - 1.67)$	$\Omega$	Li et al <sup>15</sup>	HR for incident heart failure in PLWH compared with <b>PWOH</b>
Cardiovascular events	Systematic review and meta-analysis	5	$2.36(1.50 - 3.70)$	$\Omega$	Shah et al <sup>14</sup>	HR for incident cardiovascular event (including cardiovascular mortality) in PLWH compared with PWOH

HR indicates hazard ratio; MI, myocardial infarction; PLWH, people living with HIV; PWOH, people without HIV; and SSA, sub-Saharan Africa.

**DISEASE REVIEW SERIES**

V AND CARDIOVASCULAR SEASE REVIEW SERIES

IIV AND CARDIOVASCULA ISEASE REVIEW SERIES





PLWH indicates people living with HIV.

mechanisms that underpin this inflammatory cardiomyopathy. It is likely that these mechanisms proceed at cellular and tissue levels similar to the pathophysiological mechanisms observed in other forms of viral myocarditis.41 This involves 3 distinct stages: the acute stage triggered by viral entry and replication, a subacute stage where inflammatory cells infiltrate the myocardium, and a chronic stage leading to cardiac remodeling.41 The pathological mechanisms and the evidence base underpinning them are outlined in this section and summarized in Figure 2. Few of the studies that inform these mechanisms have been performed in SSA. Table 3 summarizes some of these key studies that have specifically investigated HIV populations in SSA.

## Acute Phase of Myocardial Damage in HIV

The acute phase of myocarditis is characterized by viral entry into the cardiomyocyte and activation of the innate immune response. Historically, there has been some debate about whether HIV could directly infect cardiomyocytes. Early in the HIV epidemic, in situ hybridization of myocardial tissue from autopsies of AIDS patients with HIVAC in the United States demonstrated the presence of HIV.<sup>58-60</sup> This raised the possibility that HIV directly infected cardiomyocytes, a theory supported by in vitro experiments demonstrating that HIV can enter rat ventricular myocytes. However, these experiments also noted an absence of viral replication 3 to 4 days following infection raising questions about their mechanistic significance.<sup>61</sup> Other studies, involving human fetal cardiomyocytes, failed to demonstrate that HIV could directly infect cardiomyocytes.<sup>62</sup> In contrast, HIV is capable of infecting noncardiomyocyte inflammatory cells, specifically macrophages and T lymphocytes, within the myocardial tissue. It is from these cells that the virus is likely to exert its greatest effect on the myocardium.<sup>63,64</sup> In a study of myocardial tissue from patients with HIVAC,

only inflammatory cells and not cardiomyocytes displayed HIV genetic material.<sup>63</sup> In addition, there was perivascular macrophage infiltration of the myocardium with adjacent cardiomyocyte apoptosis.

## *HIV-Specific Proteins and Their Contribution to Myocardial Injury*

HIV envelope glycoprotein 120 (gp120), a surface protein of HIV, facilitates the virus's entry into cells.<sup>65</sup> In myocardial tissue from patients with HIVAC, gp120 was found to be expressed abundantly in macrophages and T cells but less so in cardiomyocytes.<sup>63</sup> From these infected macrophages, gp120 can induce apoptosis of adjacent cardiomyocytes via both mitochondriacontrolled and cardiomyocyte surface death receptorcontrolled pathways.61

Nef (negative regulatory factor) is an auxiliary protein of HIV that is expressed by proviral DNA.<sup>66</sup> Nef has been shown to cause direct cardiotoxicity through the inhibition of autophagy, a homeostatic cellular process that clears and degrades damaged organelles and proteins.67 Nef is detectable in the plasma of PLWH even in the presence of ART, suggesting that it may exert its effect on the myocardium in immunocompetent patients both locally or from remote tissues.<sup>68</sup> Tat (transactivator of transcription) may also contribute to myocardial injury in HIV. Targeted myocardial HIV Tat transgenic mice develop depression in both systolic and diastolic function. This appears to be mediated by cardiac mitochondrial damage indicating a possible mechanism by which Tat exerts a cardiotoxic effect.<sup>69</sup>

## *Role of Opportunistic Infections and Cardiotropic Viruses*

Both opportunistic infections and cardiotropic viruses may further exacerbate the initial myocardial insult caused by HIV. Before the widespread implementation of ART, opportunistic infections of the myocardium had been found in patients with AIDS. A study from the Congo



**Figure 2. HIV-associated nonischemic cardiomyopathy pathogenesis, the downstream clinical phenotype, and cardiovascular imaging modality.**

Inflammation stimulated by HIV infection leads to monocyte recruitment and infiltration of the myocardium with macrophages and lymphocytes. HIVspecific proteins, including HIV envelope glycoprotein 120 (gp120), Nef (negative regulatory factor), and Tat (transactivator of transcription), exert local cardiotoxic effects. Cardiotropic viruses and opportunistic infections caused by profound immunodeficiency may drive further myocardial damage. The inflammatory response, together with HIV-specific proteins, promotes fibrosis by leading to the activation of myofibroblasts and the subsequent development of myocardial fibrosis. CMR indicates cardiac magnetic resonance imaging; DCM, dilated cardiomyopathy; FDG-PET, fluorodeoxyglucose positron emission tomography; IL, interleukin; LGE, late gadolinium enhancement; and TNF-α, tumor necrosis factor α. Image credit: Sceyence Studios.

recruited 157 PLWH without cardiac disease at baseline. Over a 7-year follow-up (between 1987 and 1994), 35% developed a dilated cardiomyopathy. Autopsies were performed on 16 patients with AIDS and HIVAC, all of which demonstrated acute myocarditis with lymphocytic infiltration. Furthermore, 50% demonstrated cardiac involvement of an opportunistic infection (3 *Toxoplasma gondii*, 3 *Cryptococcus neoformans*, and 2 *Mycobacterium avium*).25

Cardiotropic viruses may also play a role in the development of nonischemic cardiomyopathy. The largest autopsy study across 440 patients with AIDS from Italy demonstrated that 30 patients had lymphocytic interstitial myocarditis. Among them, 7 had myocardial tissue culture positive for coxsackievirus, 2 with Epstein-Barr virus, and 1 with cytomegalovirus.70 A South African study performed endomyocardial biopsies in patients with HIVAC and found that almost half had myocarditis and a significant burden of cardiotropic viruses.<sup>42</sup> Tissue biopsies revealed Epstein-Barr virus (65%), herpes simplex virus (50%), parvovirus B19 (14%), and cytomegalovirus (7%). The mean viral burden in HIVAC biopsies was 2.5 viruses per case compared with 1.1 viruses per case compared with biopsies from patients with HIV-negative dilated cardiomyopathy.<sup>42</sup>

The cardiac involvement of both opportunistic infections and cardiotropic viruses has been predominantly observed in the pre-ART era. There has been a major reduction in the risk of opportunistic infections with ART use in high-income and lower-middle-income settings.<sup>71</sup> The contribution of opportunistic infections and cardiotropic viruses in the ART-era remains less certain.

## Subacute Immune Phase of Myocardial Inflammation in HIV

The HIV-infected innate immune cells and cardiac cells release cytokines and chemokines, leading to the activation and subsequent infiltration of innate immune cells

HIV AND CARDIOVASCULAR<br>DISEASE REVIEW SERIES





(*Continued* )

# **HIV AND CARDIOVASCULAR**  IIV AND CARDIOVASCULAR<br>Disease review series **DISEASE REVIEW SERIES**



 ART indicates antiretroviral therapy; BMI, body mass index; CAC, coronary artery calcium; CAD, coronary artery disease; cIMT, carotid intima-media thickness; CMR, cardiac magnetic resonance imaging; CRP, C-reactive protein; CT, computed tomography; CTCA, computed tomography coronary angiography; CVD, cardiovascular disease; CX3CR1, C-X3-C motif chemokine receptor 1; ECV, extracellular volume; FDG-PET, fluorodeoxyglucose positron emission tomography; hsCRP, high-sensitivity C-reactive protein; ICAM-1, intercellular adhesion molecule 1; IL, interleukin; LDL, low-density lipoprotein; LGE, late gadolinium enhancement; OR, odds ratio; PLWH, people living with HIV; PWOH, people without HIV; sCD163, soluble CD163; TNF-α, tumor necrosis factor α; VCAM-1, vascular cell adhesion molecule 1; and VL, viral load.

in the myocardium.72 Activated resident macrophages in particular produce proinflammatory cytokines and chemokines.<sup>73,74</sup> TNF- $\alpha$  (tumor necrosis factor  $\alpha$ ) is highly relevant and has been shown to be overexpressed in cardiac tissue of patients with HIVAC.61,75 In addition, the staining intensity of TNF- $\alpha$  in endomyocardial biopsy samples of patients with HIVAC was associated with mortality and inversely associated with CD4 T-cell counts.75 TNF- $\alpha$  levels are increased further by gp120, which can stimulate its release from activated macrophages in addition to inducing inducible nitric oxide synthase (iNOS) expression via IL (interleukin)-1β production in cardiac myocytes.<sup>76</sup> TNF- $\alpha$  impairs cardiac function by inducing cardiomyocyte apoptosis, triggering the expression of iNOS, and interfering with intracellular calcium release during systolic contraction.<sup>74</sup> To support the importance played by immune cell infiltration in HIV-induced cardiac damage, the inhibition of CCR (C-C motif chemokine receptor) 5, known to mediate both immune cell recruitment and HIV entry into cells, was found to significantly reduce viral load and prevent diastolic dysfunction in simian immunodeficiency virus-infected macaques.<sup>77</sup>

Myocardial inflammation in HIV infection can be visualized using cardiac magnetic resonance imaging (CMR). CMR utilizes T2 weighted images, native T1 mapping, and extracellular volume fraction calculation to noninvasively assess the degree of myocardial edema and diffuse myocardial fibrosis in the myocardium. A

German cohort of PLWH with well-controlled disease who underwent CMR was found to have higher levels of myocardial inflammation calculated by higher native T1 relaxation times and higher T2 signal intensity ratio.<sup>78</sup> Our analysis that pooled 7 studies from China,<sup>79</sup> Peru,<sup>80</sup> the United States, 81,82 Germany, 78 the United Kingdom, 83 and South Africa<sup>47</sup> demonstrated that PLWH had a significantly higher pooled mean difference in native T1 time compared with PWOH (33.7 ms [95% CI, 11.8– 55.7]; *P*<0.05).9 In 73 PLWH who were naive of ART in South Africa, Robbertse et al<sup>48</sup> demonstrated a higher degree of myocardial edema represented by T1 relaxation times. This is thought to represent inflammatory cell infiltration of the myocardium. In the same study, this edema appeared to resolve with significant reductions in T1 relaxation times 9 months after the initiation of ART. Finally, imaging studies evaluating the cellular infiltration phase of inflammation are absent and will likely provide supplementary information as shown with other infective states such as COVID-19.84,85

Single-cell sequencing has been extensively used to investigate CVD,<sup>86</sup> revealing changes in immune cell clusters in heart failure $87$  and MI, $88$  as well as the heterogeneity of immune cells in endarterectomy plaque samples.<sup>89</sup> Spatial transcriptomics has been used to map site-specific pathways leading to atherosclerotic plaque rupture90 and also the pathogenesis of viral myocarditis.91 Similarly, a proteomics atlas of atherosclerosis has

HIV AND CARDIOVASCULAI<br>Disease Review Series

been proposed,<sup>92</sup> and a protocol for spatial proteomics in 3-dimensional intact specimens has been developed.<sup>93</sup> However, to date, none of these techniques have been used to uncover specific immune pathways, leading to specifically HIV-driven myocardial damage. However, single-cell analysis of immune cell transcriptome during HIV infection and therapy has already been used to investigate transcriptional profiles of atypical B cells,  $94$ common and unique gene expression profiles in infected CD4+T cells,95 and novel subset of cytotoxic-like plasmacytoid dendritic cells in people with HIV-1.96 Therefore, in the near future, different omics platforms will be fundamental to better define mechanisms at the immune/ vascular and immune/cardiac interplay in PLWH.

## Cardiac Remodeling and Fibrosis Phase

Myocardial fibrosis has been increasingly identified as an important hallmark of nonischemic cardiomyopathy in HIV.9 The inflammatory processes described above, caused by HIV and triggering a cytokine storm, eventually lead to cardiomyocyte death and the replacement of myocardium with a collagen-based scar. This fibrosis can lead to impairment in contractility and function in addition to providing a proarrhythmogenic substrate increasing the risk of sudden cardiac death.<sup>97</sup> Data from the United States in 2012 demonstrated that PLWH were at a 4.5 fold higher risk of sudden cardiac death compared with those uninfected.98 A recent elegant histological study from the same group showed a 43% increase in avascular myocardial fibrosis in HIV-infected hearts compared with uninfected people on autopsy in individuals with sudden cardiac death.<sup>98</sup>

Imaging studies using CMR have demonstrated that PLWH are at a higher risk of myocardial fibrosis,

represented by the presence of late gadolinium enhancement, than PWOH.<sup>78,80,83</sup> Holloway et al<sup>7</sup> were the first to identify a high prevalence of late gadolinium enhancement in PLWH compared with PWOH in a cohort of patients from the United Kingdom (76% versus 13%). Since then, several studies have investigated the prevalence of myocardial fibrosis in PLWH using CMR, and a pooled analysis of 16 studies demonstrated a wide prevalence range of myocardial fibrosis in PLWH from 5% to 84%. Overall pooled analysis demonstrated that PLWH had over twice the risk of myocardial fibrosis, by late gadolinium enhancement, compared with uninfected comparators. The age of the population studied did not modulate this relationship, and not enough data were available to assess whether the duration of HIV infection impacted the degree of myocardial fibrosis on CMR. These findings should be interpreted with some caution due to the substantial heterogeneity and the fact that only 1 CMR study was from the SSA region (Figure 3).<sup>47</sup>

The mechanistic pathways leading to myocardial remodeling and fibrosis in HIV remain uncertain, but chronic inflammation is likely to play a crucial role.99,100 Despite HIV viral suppression, residual immune activation and subsequent chronic inflammation persist in PLWH.<sup>101</sup> Chronic HIV infection may represent a perpetuation of the classical remodeling phase of viral myocarditis. In this state of chronic immune activation, macrophages and lymphocytes that have infiltrated the myocardium are likely to be perpetually activated.<sup>102</sup> Macrophages, in particular, are central to the development of myocardial fibrosis as demonstrated in a macaque (simian immunodeficiency virus) model, where monocyte and macrophage migration to the heart was blocked using a monoclonal antibody, natalizumab. This is an IgG4κ monoclonal antibody that binds to  $\alpha$ 4-integrin and prevents inflammatory cells from



#### **Figure 3. The geographic distribution of advanced cardiovascular imaging studies in HIV.**

Numbers in parentheses represent the total number of participants with HIV included in the studies evaluated in this review. Blue indicates highincome countries, and beige indicates upper-middle–income countries classified by the World Bank Income Group.9

crossing blood vessels.<sup>103</sup> In animals that received natalizumab, there were significant reductions in CD163+ and CD68+ macrophages in the myocardium compared with controls. This was accompanied by a lower level of fibrosis.<sup>104</sup> Macrophages exhibit functional plasticity depending on their environment and can modulate their cytokine and growth factor expression profile.<sup>105</sup> It has been postulated that in a state of chronic immune activation, macrophages release profibrotic cytokines, such as IL-10 and TGF-β1 (transforming growth factor-β1), leading to increased collagen I and III deposition and subsequent myocardial fibrosis.100,102

#### *Linking Inflammation to Myocardial Fibrosis*

Attempts have been made to establish whether the myocardial fibrosis observed on CMR in PLWH is correlated with markers of inflammation. In a cohort of women living with HIV in the United States without CVD and wellcontrolled HIV, diffuse myocardial fibrosis was not only more prevalent compared with HIV-negative controls but also associated with the macrophage activation marker  $sCD163$  (soluble CD163).<sup>81</sup> This association suggests a potential role for chronic immune activation in driving the myocardial disease. However, a South African study failed to identify a link between circulating inflammatory proteins (TNFR-I [tumor necrosis factor receptor 1], IL-6, sCD14, and sCD163) and myocardial fibrosis as measured by extracellular volume fraction.49 Myocardial fibrosis has been positively associated with percentages of circulating nonclassical and intermediate monocyte phenotypes that reflect inflammation and tissue injury.49

## *Inflammatory Biomarkers and Cytokines Associated With Myocardial Fibrosis*

The cytokine TGF-β1 has been referred to as the master regulator of fibrosis<sup>106</sup> and has been implicated in HIVassociated myocardial fibrosis. TGF-β1 is elevated in PLWH and is a marker of disease progression. Levels of TGF-β1 are also increased in ART-nonadherent individuals compared with ART-adherent individuals.<sup>107,108</sup> The HIV protein Tat has also been shown to increase levels of TGFβ1. In the mouse myocardium, administration of recombinant Tat protein was found to cause myocardial fibrosis and simultaneously increase the expression of TGF- $\beta1.^{109}$ Platelet-derived TGF-β1 has also been shown to mediate HIV protease inhibitor-induced cardiac dysfunction,<sup>110</sup> highlighting the need to better understand the contribution of this protein to myocardial fibrosis in PLWH.

Two further proteins may mediate myocardial fibrosis in HIV: Gal-3 (galectin-3) and sST-2 (soluble ST-2). Gal-3 is a regulatory protein that is part of the β-galactoside and has become an important CVD biomarker. It has been implicated in a range of CVDs including heart failure,<sup>111</sup> atherosclerosis,<sup>112</sup> atrial fibrillation,<sup>113</sup> and valvular heart diseases.114 Under normal physiological conditions, it is expressed at low levels; however, in disease states such as heart failure, its expression increases substantially.115,116 It activates fibroblasts and macrophages within the myocardium, leading to the induction of myocardial fibrosis. Interestingly, the HIV protein Tat has been shown to significantly increase the expression of Gal-3. This interaction highlights a unique mechanism by which HIV infection may lead to myocardial fibrosis.117 It activates both fibroblasts and macrophages in the myocardium and has been shown to induce myocardial fibrosis.<sup>118</sup>

sST-2 is a decoy receptor for IL-33. The relationship between IL-33 and sST-2 is normally protective against myocardial fibrosis.<sup>119,120</sup> In the context of heart failure, sST-2 serves as a biomarker of myocardial fibrosis and cardiac remodeling. sST-2 levels correlate with the severity and prognosis of heart failure, making it a valuable tool for risk stratification and monitoring of disease progression.<sup>121</sup> Secemsky et al<sup>122</sup> demonstrated in PLWH a relationship between diastolic dysfunction, sST-2, and all-cause mortality. They suggest that sST-2 in PLWH may be a mediator of the myocardial fibrosis extensively described in CMR studies. A direct link between myocardial fibrosis and inflammation was offered by 2 recent studies performed in women with HIV, demonstrating, respectively, a correlation between plasma osteopontin, a predictor of heart failure, myocardial fibrosis and the expression of 2 important chemokine receptors central to myocardial inflammation, CCR2 and CX3CR1 (C-X3-C motif chemokine receptor 1) in monocytes,<sup>123</sup> and a correlation between myocardial fibrosis, reduced diastolic function, and elevated systemic monocyte activation.<sup>81</sup>

While our understanding of myocardial inflammation in HIV remains incomplete, the increased circulating levels of Gal-3 and sST-2 in PLWH highlight potential pathways through which HIV may contribute to myocardial fibrosis, suggesting new avenues for targeted therapeutic interventions and research.

## MECHANISMS OF CORONARY ARTERY DISEASE IN HIV INFECTION

#### Vascular Inflammation and Atherosclerosis

It is well established that atherosclerosis, the process of plaque formation in the vascular intima, is a chronic vascular inflammatory process.<sup>124,125</sup> One of the key mechanisms postulated in HIV-associated CVD is that the virus causes vascular inflammation, leading to atherosclerosis and subsequent ischemic heart disease.<sup>6,126</sup> Molecular and anatomic imaging alongside immunologic studies provide insights into the pathogenesis of atherosclerosis in HIV. The hypothesized pathological mechanisms of HIV-associated coronary artery disease are outlined in Figure 4.

#### *Molecular Imaging*

Vascular 18F-fluorodeoxyglucose positron emission tomography (18F-FDG-PET) imaging identifies areas



**Figure 4. Mechanisms of HIV-associated atherosclerotic coronary artery disease pathogenesis, clinical phenotype, and the appropriate cardiovascular imaging modality.**

HIV infection causes a strong inflammatory response, which leads to the release of cytokines that cause monocyte activation and recruitment to the endothelium. Adhesion molecules are also stimulated, which facilitates macrophages entering the endothelial layer. Specific HIV proteins drive endothelial dysfunction, stimulate adhesion molecules, and can impact the efflux of cholesterol in macrophages contributing to foam cell formation. CCL2 indicates C-C motif chemokine ligand 2; CD14, cluster of differentiation 14; CMR, cardiac magnetic resonance imaging; CRP, C-reactive protein; CTCA, computed tomography coronary angiography; FDG-PET, fluorodeoxyglucose positron emission tomography; IL, interleukin; LGE, late gadolinium enhancement; Nef, negative regulatory factor; sCD163, soluble CD163; Tat, transactivator of transcription; and TNF-α, tumor necrosis factor α. Image credit: Sceyence Studios.

of high glucose uptake by activated macrophages within atherosclerotic plaques. It has been used in HIV-negative populations to identify and characterize vascular inflammation in atherosclerosis.<sup>127</sup> Higher 18F-FDG uptake indicates increased inflammatory activity and faster progression of atherosclerosis, which is associated with an increased risk of future cardiovascular events.128,129

Subramanian et al<sup>8</sup> first demonstrated that  $PLWH$ recruited from North America, who were on stable ART and without CVD, had higher aortic 18F-FDG uptake compared with PWOH matched for baseline cardiovascular risk. This was supported by 2 further studies across PLWH without CVD and on ART.<sup>51,130</sup> Lawal et al<sup>51</sup> compared the aortic tissue background ratio (TBR) of PLWH in South Africa to age- and sex-matched controls and found marginally higher levels of vascular inflammation that appeared independent of CD4 T-cell count and HIV viral load. However, the retrospective study design and

the fact that participants were undergoing 18F-FDG-PET in view of suspected inflammatory disease or malignancy limit the validity of these findings.

Other studies using 18F-FDG-PET in American and Danish cohorts of PLWH to assess vascular inflammation have found no difference in aortic FDG uptake between PLWH and PWOH.<sup>131,132</sup> One clinical trial conducted in the United States investigated the role of statins in modulating vascular inflammation in ART-established PLWH but without CVD.<sup>133</sup> At 12 months, there was no difference in aortic 18F-FDG uptake in those receiving statins compared with placebo.

Importantly, 18F-FDG-PET studies to date have been restricted to predominantly male participants in their fifth decade in nonendemic regions usually with known cardiovascular risk factors.<sup>9</sup> In contrast, the majority of PLWH in SSA are women, typically younger, and with a lower cardiovascular risk profile.<sup>28,53,134,135</sup> There exists

only 1 molecular imaging study evaluating vascular inflammation in HIV populations from SSA.<sup>51</sup>

## *Evidence From Anatomic Imaging*

Computed tomography coronary angiography (CTCA) has been used to further describe the extent and severity of coronary artery disease in HIV. Again, the majority of studies originate from North America/Europe and have shown diverging results. A cohort of North American homosexual men with HIV and high prevalence of vascular risk factors showed that 8 out of 10 individuals had evidence of coronary plaque and were at a 1.5-fold higher risk of coronary disease compared with uninfected individuals.<sup>136</sup> Later studies from Canada,<sup>137</sup> Switzerland,<sup>138</sup> and the United States,<sup>139</sup> however, failed to detect an association between HIV infection and the presence of coronary plaque in PLWH. Studies have also indicated a heightened risk of obstructive and clinically significant coronary stenosis, defined as a  $>50\%$  obstruction of the coronary lumen, in PLWH.<sup>136,140</sup>

Evidence synthesis of CTCA studies showed significant heterogeneity, with the prevalence of coronary artery disease in HIV ranging from 17.1% to 77.8%.<sup>9</sup> Similarly, compared with uninfected comparators, pooled analysis showed significant heterogeneity with prevalence ratios ranging from 0.33 (95% CI, 0.01-15.90) to 5.19 (95% Cl, 1.26-21.42).<sup>9</sup> Metaregressions to explore whether particular HIV-specific or non-HIV risk factors modulated this relationship indicated that only the proportion of smokers in the population and mean body mass index were significantly associated with coronary artery stenosis on CTCA. There was no relationship between mean CD4 T-cell count, nadir CD4 T-cell count, time on ART, and the proportion of patients on ART.

There is little evidence from SSA to support a relationship between HIV and coronary artery disease in PLWH. Alencherry et  $a^{145}$  compared the coronary artery calcium score of PLWH on ART to age- and sex-matched and risk factor–matched PWOH across populations in Uganda and the United States. Ugandans had markedly lower coronary artery calcium scores than individuals from the United States. Furthermore, HIV was not associated with a higher coronary artery calcium score.<sup>45</sup> Recent data from Uganda showed a near 2-fold lower prevalence of coronary pathology across PLWH compared with uninfected controls alongside lower rates of coronary calcification in HIV.<sup>50</sup> Carotid intimal-media thickness (cIMT) measurement using ultrasound is a cheaper alternative assessment of atherosclerosis to CTCA. Studies in Uganda and South Africa have failed to show a difference in cIMT between PLWH compared with HIV-negative comparators.<sup>43,141</sup> The largest study of its kind including 8872 participants from Kenya, Burkina Faso, Ghana, and South Africa did not find HIV to be a risk factor for higher cIMT.46

## *Immunologic Evidence*

The immunologic evidence for HIV-associated atherosclerosis is in part derived from several inflammatory biomarker studies. Circulating inflammatory biomarkers strongly associated with atherosclerosis such as IL-6 and C-reactive protein, and levels of the coagulation marker D-dimer, are consistently elevated in PLWH even in the presence of controlled viremia.<sup>142,143</sup> This was demonstrated in the SMART study (Strategies for Management of Antiretroviral Therapy) that revealed increased mortality in participants with higher IL-6 and D-dimer levels, a finding reproduced by others.<sup>144,145</sup> More recently, several soluble markers of inflammation (sCD14, sCD163, TNFR-I, TNFR-II, CCL [C-C motif chemokine ligand] 5, CX3CL1 [C-X3-C motif chemokine ligand 1], and IL-10) have shown associations with pericoronary inflammation in HIV.146 Similarly, within HIV-endemic areas in SSA, particularly in Uganda, there was a clear association between inflammation and subclinical CAD in adult women, both with and without HIV.<sup>57</sup> hsCRP (highsensitivity C-reactive protein) showed a positive correlation with traditional cardiometabolic risk factors in PLWH.52 Moreover, persistent immune activation despite viral suppression mediated by ART was predictive of future atherosclerotic burden among PLWH.<sup>53</sup> Altered lipid profiles and inflammation were also linked to vascular disease in HIV-positive children.<sup>54</sup> Additionally, in newly diagnosed, untreated HIV-1-infected South Africans, systemic inflammation was associated with endothelial dysfunction, a significant contributor to atherosclerosis development.43 Overall, these findings suggest that inflammatory biomarkers that are known to predict cardiovascular events in the general population and are associated with major adverse cardiovascular events are both elevated and predict cardiovascular events in PLWH.147

It is noteworthy that among PLWH in Uganda, there is an expansion of the inflammatory monocyte subset, accompanied by enhanced expression of the monocyte CX3CR1 chemokine receptor.<sup>56</sup> This finding holds significance in the context of CVD, considering that monocytes from PLWH on ART display a high frequency of cells producing IL-1 $\beta$ .<sup>148</sup> These cells have been shown to be a major source of IL-6 production and systemic inflammation in PLWH at risk for CVD. Pharmacological targeting of IL-1β has been recently shown to significantly reduce the rate of recurrent cardiovascular events.<sup>149</sup> Similarly, the targeting of IL-6 is currently under intense investigation in people with CVDs (<https://www.clinicaltrials.gov>; unique identifier: NCT05021835).<sup>150</sup> Other proinflammatory cytokines, such as IL-17A, macrophage inflammatory protein-1 alpha (MIP-1 $\alpha$ /CCL3), IL-4, and type I interferon, have also been found to be associated with subclinical atherosclerosis in PLWH with suppressed viremia.151,152

The above studies suggest an inflammatory process mediates the risk of coronary atherosclerosis in HIV both

HIV AND CARDIOVASCULAI<br>Disease Review Series

in the Global North and the Global South. HIV infection leads to an increase in IL-6 and TNF- $\alpha$  secretion that stimulate the release of chemokine (C-C motif) ligand 2 (CCL2 also known as monocyte chemoattractant protein-1), which is a key activator and recruiter of monocytes to the vascular endothelium.153,154 Several studies have established a connection between CCL2 and HIVinduced atherosclerosis. CCL2 is found in high levels in PLWH and is associated with immunodeficiency (high viral load and low CD4 T-cell counts) and atherosclerotic burden in the thoracic aorta.<sup>155</sup> In a cohort of men with HIV, CCL2 levels were higher compared with HIV-negative controls and showed a significant association with subclinical atherosclerosis and coronary stenosis detected by CTCA. Besides CCL2, several other markers of macrophage activation including galectin-3-binding protein, sCD163, and soluble CD14 (cluster of differentiation 14) have been detected at increased levels in PLWH and associated with coronary atherosclerosis.156,157

This immune response to HIV infection stimulates endothelial cells to recruit leukocytes to the subendothelial layer and in the perivascular space. During HIV infection, elevated levels of cytokines (TNF- $\alpha$ , IL-1, and IL-4) and chemokines (CCL2 and CCL3) upregulate adhesion molecules that recruit monocytes to the endothelium.<sup>158</sup> These monocytes, driven by the inflammatory milieu, subsequently differentiate into macrophages and foam cells resulting in the initiation and acceleration of atherosclerosis. Evidence for this hypothesis is shown by the efficacy of maraviroc, a CCR5 antagonist, which led to significant improvements in several markers for cardiovascular risk, endothelial dysfunction, arterial stiffness, and early carotid atherosclerosis in HIV-treated patients<sup>159</sup> and decreased ritonavir-induced atherogenesis and advanced atherosclerosis by reducing vascular inflammation in mice.160

Two HIV proteins, Tat and Nef, have also been implicated in the pathogenesis of HIV-associated atherosclerosis. Tat, whose primary role in HIV infection is to improve the efficiency of viral replication, can be excreted by infected T cells and monocytes into the extracellular environment. Its implication in atherosclerosis is via several postulated mechanisms. First, it promotes cytokine production from monocytes and stimulates the release of CCL2.161 Second, it increases the adhesion of monocytes and T cells to the vascular endothelium.162 Third, it can induce endothelial dysfunction<sup>163</sup> and, in mouse models, can directly increase the size of atherosclerotic lesions.<sup>164</sup> Nef protein also drives lymphocyte chemotaxis and macrophage activation via CD36.165 In addition, through the inhibition of ATP-binding cassette transporter A1, it impairs cholesterol efflux from infected macrophages and facilitates the transformation of these into foam cells.166

More recently, clonal hematopoiesis was found to be higher in PLWH, associated with subclinical atherosclerosis

but not with elevated levels of inflammatory biomarkers.<sup>167</sup> Immune cell senescence has also been shown to predict subclinical atherosclerosis in PLWH.<sup>168,169</sup> There is also evidence that HIV alters the metabolism in immune cells leading to a proinflammatory phenotype.170 Finally, the potential contribution of trained immunity to HIVassociated atherosclerosis should also be taken into consideration. Following an initial infection, long-term epigenetic reprogramming allows innate immune cells to provide enhanced response to subsequent infections. Although only a few studies have focused on trained immunity in the context of HIV infection,<sup>171</sup> trained immunity has received growing attention in atherosclerosis<sup>172</sup> and may represent an important missing link to explain HIV-associated cardiovascular comorbidities.

In summary, the mechanisms through which HIV drives atherosclerosis are related predominantly to a surge of cytokines and chemokines during HIV infection accompanied by the atherogenic effects of HIV-specific proteins. This leads to systemic immune cell activation, accelerated immune cell senescence, and ultimately increased endothelial dysfunction and vascular inflammation. Of note, the use of statins such as rosuvastatin reduced significantly several inflammatory biomarkers and immune activation in ART-treated individuals.173 Further aspects such as the contribution of immunometabolism or trained immunity to HIV-associated CVD still need to be fully elucidated.

## Role of ART in Cardiovascular Pathology

## *Class of ART and Cardiovascular Pathology*

ART has transformed the HIV epidemic and changed the nature of cardiovascular risk in PLWH. With earlier ART regimens, concerns were raised about the cardiotoxic effects. The DAD study (Data Collection on Adverse Events of Anti-HIV Drugs) provided evidence across a cohort of over 23 000 patients recruited from Europe and Australia between 1999 and 2001, showing that the use of ART was associated with a 26% relative increase in the rate of MI during the first 6 years of treatment.<sup>174</sup> The DAD study was then followed by 2 clinical trials evaluating intermittent ART and delayed ART initiation and risk of adverse events. The SMART study randomized patients recruited from the United Kingdom, Australia, and Denmark with a CD4 T-cell count over 350 cells/μL to receive either continuous ART compared with episodic ART when the CD4 count dropped below 250 cells/μL. Continuous ART provided a 70% reduction in the risk of major cardiovascular events.175 Notably, major adverse cardiovascular events occurred 5× more often than opportunistic infections in the treatment interruption group.175 The International Network for Strategic Initiatives in Global HIV Trials - Strategic Timing of Antiretroviral Therapy study (INSIGHT-START) was a large multinational trial that recruited 4685 PLWH across 35

countries including Uganda, Mali, Nigeria, and South Africa. Delaying ART in asymptomatic immunocompetent PLWH in the trial was associated with a high risk of both AIDS and non-AIDS events.176

Protease inhibitors (PI) have traditionally been thought to increase the risk of CVD. Further analyses from the DAD cohort demonstrated an increased risk of MI with the use of protease inhibitors. This risk appeared to be at least partly explained by dyslipidemia.<sup>177</sup> A meta-analysis of observational studies indicated that recent use of PI was associated with a doubling in the odds of MI.<sup>178</sup> The authors highlight the low quality of the studies that inform this evidence. A meta-analysis of randomized trials, underpowered to assess cardiovascular outcomes and of short duration, found no association between PI use and MI.<sup>179</sup> The cardiovascular risk is not equal across different PIs. The DAD study demonstrated that the cumulative use of ritonavir-boosted darunavir, but not ritonavir-boosted atazanavir, was associated with an increased risk of CVD.180 Although the increased risk of CVD with PIs is likely a class effect, atazanavir appears not to confer an excess risk of CVD.6,181

Abacavir, a nucleoside reverse transcriptase inhibitor, has been the focus of significant attention due to concerns about its CVD risk profile. The DAD cohort identified that recent abacavir use was associated with an increased risk of MI and a meta-analysis of observational studies confirmed this association.<sup>178,182</sup> How abacavir exerts detrimental cardiovascular effects is not known but may involve platelet activation<sup>183,184</sup> and endothelial dysfunction.185 The cardiotoxicity of abacavir remains an area of uncertainty and debate in light of a US Food and Drug Administration sponsored trial meta-analysis of randomized control trials, which did not show an excess of MI.<sup>186</sup>

The relatively newer integrase strand transfer inhibitors (INSTIs) are now the mainstay of ART and recommended first-line treatment in HIV guidelines, given their improved virological efficacy and lower potential for drug-drug interactions. INSTIs, although initially thought to be less cardiotoxic, have been found to increase the incidence of arterial hypertension and diabetes.<sup>187</sup> The International Cohort Consortium of Infectious Disease (RESPOND) collaboration (comprising 17 European and Australian HIV cohorts) also found an increased risk of cardiovascular events with the use of INSTIs that was observed only within the first 2 years of use.<sup>188</sup> In contrast, the HIV Swiss cohort did not find the same risk.<sup>189</sup> INSTIs are still considered less cardiotoxic; however, the excess risk detected in the RESPOND collaboration may represent a bias toward prescribing INSTI-based regimens to patients at higher risk of CVD.

## *How ART Modulates HIV-Associated CVD Pathogenesis: Imaging Data*

Advanced cardiovascular imaging has provided a greater understanding of how ART treatment affects

the progression of HIV-associated CVD. The degree to which myocardial inflammation occurs before ART initiation has been assessed by CMR in several studies. Menacho et al<sup>80</sup> recruited 51 PLWH in Peru, 25 of whom were treatment-naive. Untreated PLWH had high T1 values and extracellular volume compared with those on ART, suggesting a greater degree of myocardial edema and fibrosis before initiating ART.80

Two studies evaluated the changes in myocardial pathology with ART initiation showing diverging results. One case series (n=17) from Mexico, involving ART-naive patients, showed that 1 in 3 individuals had evidence of myocarditis on CMR. Following ART establishment, myocarditis resolved in 4 of the patients but persisted in 2, and developed in a further 3 patients. While these findings may hint toward the possibility of an immune reconstitution syndrome propagating myocardial inflammation, others have demonstrated the protective effect of ART.<sup>190</sup> Robbertse et al<sup>48</sup> demonstrated a reduction of magnetic resonance imaging–based parameters of myocardial fibrosis (global native T1 times and extracellular volume fraction) following ART initiation with a concurrent improvement in CD4 counts and viral load suppression. Although this study did not assess clinical outcomes, measure myocardial inflammatory cell infiltration, or assess correlations with circulating markers of cardiac injury, it demonstrates 2 important points. First, untreated HIV causes myocardial injury based on imaging. This is likely driven by the cytokines released during HIV replication. Separate biomarker studies have supported these radiological findings, showing elevated troponin levels in PLWH who are ART naive and abating with viral suppression.<sup>191</sup> Second, these findings highlight ART's potential cardioprotective role in reversing inflammatory changes caused by HIV.

The impact of ART on vascular inflammation was explored by Zanni et al<sup>192</sup> in 12 ART-naive men who underwent an 18F-FDG-PET scan and CTCA before and 6 months after the initiation of ART. Inflammation in the aorta increased marginally 6 months after the initiation of ART despite a reduction in viral load and improved immune function. The total coronary plaque volumes also increased in those who had any coronary plaque on CTCA at baseline (although the study was not powered for this end point). Ongoing vascular inflammation after the initiation of ART may represent ongoing immune activation, ART-mediated effects through changes in cholesterol levels, or a degree of immune reconstitution syndrome. To observe a reduction in vascular inflammation, a longer delay between initiating ART and performing the PET scan may be necessary.

Both the SMART and INSIGHT-START trials provide robust evidence of the importance of continual and prompt ART treatment.<sup>175,176</sup> Across patients on already established ART, it may be important that maintaining viral suppression and preventing transient HIV viremia



**Figure 5. Approach to studying HIV-associated cardiovascular disease.**

The imaging modalities highlighted are for research purposes and are not clinical recommendations. CTCA indicates computed tomography coronary angiography; FDG-PET, fluorodeoxyglucose positron emission tomography; MRI, magnetic resonance imaging; and PBMC, peripheral blood mononuclear cells. Image credit: Sceyence Studios.

may be an important strategy to avert future CVD. Several mechanistic imaging studies support the hypothesis that consistent viral load suppression reduces cardiovascular risk. McLaughlin et al<sup>193</sup> recruited 152 PLWH with suppressed viral loads and demonstrated that higher levels of HIV RNA and HIV DNA were associated with incident plaque development after adjusting for cardiovascular and HIV-associated risk factors. This is supported by The Study to Understand the Natural History of HIV and AIDS in the Era of Effective Therapy (SUN Study) that demonstrated among participants on ART, suppressed viral load at baseline, and maintaining that suppression during the median 2-year follow-up was associated with less cIMT progression.194 Additionally, progression of coronary artery stenosis is greater in patients who have suboptimal ART adherence and have blips in their viral load (measured as 50–500 copies/mL) over a median of 4.5 years of follow-up.195 Viral blips can occur as a consequence of a brief increase in replication from the HIV reservoir. This reservoir consists of long-lived cells harboring HIV in a latent state with the potential to replicate.196 Further research is needed to better understand the significance of viral blips, especially as there are marked disparities between populations who are adherent to ART and those who are not. Despite this evidence, however, the degree to which low levels of detectable viral load in PLWH on ART increase the risk of CVD is not fully understood.

## FUTURE DIRECTIONS

Inflammation and immune activation persist even in patients on ART with viral suppression. The mechanisms

by which this leads to HIV-associated CVD remain unknown. The current understanding of the mechanisms and timing of myocardial inflammation and fibrosis development in PLWH is limited. The contribution of chronic inflammation to myocardial fibrosis prevalence and the subsequent impact on CVD outcomes remains unknown. As such, despite the use of ART, PLWH continue to experience an excess risk of CVD. Increasing evidence hints at the importance of a nonischemic cardiomyopathic process driving HIV-associated CVD, particularly in HIV-endemic regions.

The REPRIEVE trial underscores the necessity for primary prevention strategies in HIV-associated CVD but also demonstrates that future research must rigorously address the notable underrepresentation of SSA populations.<sup>11,35</sup> Carefully designed studies across appropriate populations are now urgently needed to define the primary pathogenic mechanisms explaining the rising epidemic of HIV-associated CVD (Figure 5). How these pathogenic mechanisms manifest in anatomic, functional, and molecular cardiovascular imaging and relate to the underlying immune basis propagating HIV-associated CVD is likely to be a key area of research in the near future.

## ARTICLE INFORMATION

#### Affiliations

Kings College London BHF Centre, School of Cardiovascular and Metabolic Medicine & Sciences, United Kingdom (J.A.H.). Department of Clinical Research (R.A.F.) and Department of Non-Communicable Disease Epidemiology (A.S.V.S.), London School of Hygiene and Tropical Medicine, United Kingdom. Biomedical Research and Training Institute, Harare, Zimbabwe (R.A.F.). Department of Radiology, Aga Khan University Nairobi, Kenya (S.N.G.). Department of Medical Microbiology and Immunology, Faculty of Health Sciences (M.W.M.) and Kenya AIDS Vaccine Initiative (KAVI)-Institute of Clinical Research (M.W.M.), University of Nairobi, Kenya. School of Infection and Immunity, College of Medical, Veterinary and Life Sciences, University of Glasgow, United Kingdom (P.M.). Department of Pharmacy, School of Medicine and Surgery, University of Naples Federico II, Italy (P.M.). Africa-Europe Cluster of Research Excellence in Non-Communicable Diseases and Multimorbidity, African Research Universities Alliance and The Guild of European Research-Intensive Universities, Glasgow, United Kingdom (P.M.). Department of Cardiology, North Bristol NHS Trust, United Kingdom (S.R.A.). Department of Cardiology, Imperial College NHS Trust, London, United Kingdom (A.S.V.S.).

#### Sources of Funding

J.A. Hudson was funded by the National Institute for Health and Research (NIHR; NIHR Academic Clinical Fellowship grant ACF-2021-17-001).

#### **Disclosures**

None.

#### REFERENCES

- 1. Trickey A, May MT, Vehreschild J, Obel N, Gill MJ, Crane H, Boesecke C, Samji H, Grabar S, Cazanave C, et al; Antiretroviral Therapy Cohort Collaboration (ART-CC). Cause-specific mortality in HIV-positive patients who survived ten years after starting antiretroviral therapy. *PLoS One*. 2016;11:e0160460. doi: 10.1371/journal.pone.0160460
- 2. Smith C, Sabin CA, Lundgren JD, Thiebaut R, Weber R, Law M, Monforte A, Kirk O, Friis-Moller N, Phillips A, et al; Data Collection on Adverse Events of Anti-HIV drugs (D:A:D) Study Group. Factors associated with specific causes of death amongst HIV-positive individuals in the D:A:D study. *AIDS*. 2010;24:1537–1548. doi: 10.1097/QAD.0b013e32833a0918
- 3. Mocroft A, Reiss P, Gasiorowski J, Ledergerber B, Kowalska J, Chiesi A, Gatell J, Rakhmanova A, Johnson M, Kirk O, et al; EuroSIDA Study Group. Serious fatal and nonfatal non-AIDS-defining illnesses in Europe. *J Acquir Immune Defic Syndr*. 2010;55:262–270. doi: 10.1097/QAI.0b013e3181e9be6b
- 4. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. *J Clin Endocrinol Metab*. 2007;92:2506– 2512. doi: 10.1210/jc.2006-2190
- 5. McGettrick P, Barco EA, Mallon PWG. Ageing with HIV. *Healthcare (Basel)*. 2018;6:17. doi: 10.3390/healthcare6010017
- 6. Feinstein MJ, Hsue PY, Benjamin LA, Bloomfield GS, Currier JS, Freiberg MS, Grinspoon SK, Levin J, Longenecker CT, Post WS. Characteristics, prevention, and management of cardiovascular disease in people living with HIV: a scientific statement from the American Heart Association. *Circulation*. 2019;140:e98–e124. doi: 10.1161/CIR.0000000000000695
- 7. Holloway CJ, Ntusi N, Suttie J, Mahmod M, Wainwright E, Clutton G, Hancock G, Beak P, Tajar A, Piechnik SK, et al. Comprehensive cardiac magnetic resonance imaging and spectroscopy reveal a high burden of myocardial disease in HIV patients. *Circulation*. 2013;128:814–822. doi: 10.1161/CIRCULATIONAHA.113.001719
- 8. Subramanian S, Tawakol A, Burdo TH, Abbara S, Wei J, Vijayakumar J, Corsini E, Abdelbaky A, Zanni MV, Hoffmann U, et al. Arterial inflammation in patients with HIV. *JAMA*. 2012;308:379–386. doi: 10.1001/jama.2012.6698
- 9. Hudson JA, Majonga ED, Ferrand RA, Perel P, Alam SR, Shah ASV. Association of HIV infection with cardiovascular pathology based on advanced cardiovascular imaging: a systematic review. *JAMA*. 2022;328:951–962. doi: 10.1001/jama.2022.15078
- 10. Soares C, Samara A, Yuyun MF, Echouffo-Tcheugui JB, Masri A, Samara A, Morrison AR, Lin N, Wu WC, Erqou S. Coronary artery calcification and plaque characteristics in people living with HIV: a systematic review and meta-analysis. *J Am Heart Assoc*. 2021;10:e019291. doi: 10.1161/JAHA.120.019291
- 11. Grinspoon SK, Fitch KV, Zanni MV, Fichtenbaum CJ, Umbleja T, Aberg JA, Overton ET, Malvestutto CD, Bloomfield GS, Currier JS, et al; REPRIEVE Investigators. Pitavastatin to prevent cardiovascular disease in HIV infection. *N Engl J Med*. 2023;389:687–699. doi: 10.1056/NEJMoa2304146
- 12. Rerkpattanapipat P, Wongpraparut N, Jacobs LE, Kotler MN. Cardiac manifestations of acquired immunodeficiency syndrome. *Arch Intern Med*. 2000;160:602–608. doi: 10.1001/archinte.160.5.602
- 13. Felker GM, Thompson RE, Hare JM, Hruban RH, Clemetson DE, Howard DL, Baughman KL, Kasper EK. Underlying causes and long-term survival in patients with initially unexplained cardiomyopathy. *N Engl J Med*. 2000;342:1077–1084. doi: 10.1056/NEJM200004133421502
- 14. Shah ASV, Stelzle D, Lee KK, Beck EJ, Alam S, Clifford S, Longenecker CT, Strachan F, Bagchi S, Whiteley W, et al. Global burden

of atherosclerotic cardiovascular disease in people living with HIV: systematic review and meta-analysis. *Circulation*. 2018;138:1100–1112. doi: 10.1161/CIRCULATIONAHA.117.033369

- 15. Li X. HIV infection and risk of heart failure: a meta-analysis and systematic review. *Acta Biochim Pol*. 2022;69:531–535. doi: 10.18388/abp.2020\_5718
- 16. Yen YF, Ko MC, Yen MY, Hu BS, Wang TH, Chuang PH, Lai HH, Chen CC, Deng CY. Human immunodeficiency virus increases the risk of incident heart failure. *J Acquir Immune Defic Syndr*. 2019;80:255–263. doi: 10.1097/QAI.0000000000001917
- 17. Lumsden RH, Bloomfield GS. The causes of HIV-associated cardiomyopathy: a tale of two worlds. *Biomed Res Int*. 2016;2016:1–9. doi: 10.1155/2016/8196560
- 18. Bijl M, Dieleman JP, Simoons M, van der Ende ME. Low prevalence of cardiac abnormalities in an HIV-seropositive population on antiretroviral combination therapy. *J Acquir Immune Defic Syndr*. 2001;27:318–320. doi: 10.1097/00126334-200107010-00018
- 19. Erqou S, Lodebo BT, Masri A, Altibi AM, Echouffo-Tcheugui JB, Dzudie A, Ataklte F, Choudhary G, Bloomfield GS, Wu WC, et al. Cardiac dysfunction among people living with HIV. *JACC Heart Fail*. 2019;7:98–108. doi: 10.1016/j.jchf.2018.10.006
- 20. Kofi Owusu I. Echocardiographic abnormalities in patients with HIV infection at Komfo Anokye teaching hospital, Ghana. *J Gen Pract*. 2014;2:1000148. doi: 10.4172/2329-9126.1000148
- 21. Olusegun-Joseph DA, Ajuluchukwu JNA, Okany CC, Mbakwem AC, Oke DA, Okubadejo NU. Echocardiographic patterns in treatment-naïve HIV-positive patients in Lagos, south-west Nigeria. *Cardiovasc J Afr*. 2012;23:e1–e6. doi: 10.5830/CVJA-2012-048
- 22. Hamadou B, Ngweth MN, Fotso MM, Mfeukeu-Kuate L, Jingi AM, Noubiap JJ, Nganou CN, Boombhi J, Amougou SN, Menanga A, et al. Echocardiographic and electrocardiographic abnormalities in adults living with human immunodeficiency virus: a cross-sectional study in the Yaoundé Central Hospital, Cameroon. *Cardiovasc Diagn Ther*. 2017;7:607–615. doi: 10.21037/cdt.2017.10.12
- 23. Hakim JG, Matenga JA, Siziya S. Myocardial dysfunction in human immunodeficiency virus infection: an echocardiographic study of 157 patients in hospital in Zimbabwe. *Heart*. 1996;76:161–165. doi: 10.1136/hrt.76.2.161
- 24. Feinstein MJ, Steverson AB, Ning H, Pawlowski AE, Schneider D, Ahmad FS, Sanders JM, Sinha A, Nance RM, Achenbach CJ, et al. Adjudicated heart failure in HIV-infected and uninfected men and women. *J Am Heart Assoc*. 2018;7:e009985. doi: 10.1161/JAHA.118.009985
- 25. Longo-Mbenza B, Seghers KV, Phuati M, Bikangi FN, Mubagwa K. Heart involvement and HIV infection in African patients: determinants of survival. *Int J Cardiol*. 1998;64:63–73. doi: 10.1016/s0167-5273(97)00321-5
- 26. Twagirumukiza M, Nkeramihigo E, Seminega B, Gasakure E, Boccara F, Barbaro G. Prevalence of dilated cardiomyopathy in HIV-infected African patients not receiving HAART: a multicenter, observational, prospective, cohort study in Rwanda. *Curr HIV Res*. 2007;5:129–137. doi: 10.2174/157016207779316288
- 27. Sliwa K, Carrington MJ, Becker A, Thienemann F, Ntsekhe M, Stewart S. Contribution of the human immunodeficiency virus/acquired immunodeficiency syndrome epidemic to de novo presentations of heart disease in the Heart of Soweto Study cohort. *Eur Heart J*. 2012;33:866–874. doi: 10.1093/eurheartj/ehr398
- 28. UNAIDS. 2023 UNAIDS Global AIDS Update. Joint United Nations Programme on HIV/AIDS (UNAIDS); 2023. [https://thepath.unaids.org/wp](https://thepath.unaids.org/wp-content/themes/unaids2023/assets/files/2023_report.pdf)[content/themes/unaids2023/assets/files/2023\\_report.pdf](https://thepath.unaids.org/wp-content/themes/unaids2023/assets/files/2023_report.pdf)
- 29. Freiberg MS, Chang CCH, Kuller LH, Skanderson M, Lowy E, Kraemer KL, Butt AA, Bidwell Goetz M, Leaf D, Oursler KA, et al. HIV infection and the risk of acute myocardial infarction. *JAMA Intern Med*. 2013;173:614–622. doi: 10.1001/jamainternmed.2013.3728
- 30. Drozd DR, Kitahata MM, Althoff KN, Zhang J, Gange SJ, Napravnik S, Burkholder GA, Mathews WC, Silverberg MJ, Sterling TR, et al. Increased risk of myocardial infarction in HIV-infected individuals in North America compared to the general population. *J Acquir Immune Defic Syndr*. 2017;75:568–576. doi: 10.1097/QAI.0000000000001450
- Johnston PI, Wright SW, Orr M, Pearce FA, Stevens JW, Hubbard RB, Collini PJ. Worldwide relative smoking prevalence among people living with and without HIV. *AIDS*. 2021;35:957–970. doi: 10.1097/QAD.0000000000002815
- 32. Davis K, Perez-Guzman P, Hoyer A, Brinks R, Gregg E, Althoff KN, Justice AC, Reiss P, Gregson S, Smit M. Association between HIV infection and hypertension: a global systematic review and meta-analysis of cross-sectional studies. *BMC Med*. 2021;19:105. doi: 10.1186/s12916-021-01978-7
- 33. Clark SJ, Gómez-Olivé FX, Houle B, Thorogood M, Klipstein-Grobusch K, Angotti N, Kabudula C, Williams J, Menken J, Tollman S. Cardiometabolic

disease risk and HIV status in rural South Africa: establishing a baseline. *BMC Public Health*. 2015;15:135. doi: 10.1186/s12889-015-1467-1

- 34. Brown TT, Cole SR, Li X, Kingsley LA, Palella FJ, Riddler SA, Visscher BR, Margolick JB, Dobs AS. Antiretroviral therapy and the prevalence and incidence of diabetes mellitus in the multicenter AIDS cohort study. *Arch Intern Med*. 2005;165:1179–1184. doi: 10.1001/archinte.165.10.1179
- 35. Alam SR, Mureithi M, Ferrand R. Pitavastatin and cardiovascular disease in HIV. *N Engl J Med*. 2023;389:e46. doi: 10.1056/NEJMc2311117
- 36. Hyle EP, Mayosi BM, Middelkoop K, Mosepele M, Martey EB, Walensky RP, Bekker LG, Triant VA. The association between HIV and atherosclerotic cardiovascular disease in sub-Saharan Africa: a systematic review. *BMC Public Health*. 2017;17:954. doi: 10.1186/s12889-017-4940-1
- 37. Yao H, Ekou A, Niamkey T, Hounhoui Gan S, Kouamé I, Afassinou Y, Ehouman E, Touré C, Zeller M, Cottin Y, et al. Acute coronary syndromes in sub-Saharan Africa: a 10-year systematic review. *J Am Heart Assoc*. 2022;11:e021107. doi: 10.1161/JAHA.120.021107
- 38. Hertz JT, Reardon JM, Rodrigues CG, De Andrade L, Limkakeng AT, Bloomfield GS, Lynch CA. Acute myocardial infarction in sub-Saharan Africa: the need for data. *PLoS One*. 2014;9:e96688. doi: 10.1371/journal.pone.0096688
- 39. Sliwa K, Wilkinson D, Hansen C, Ntyintyane L, Tibazarwa K, Becker A, Stewart S. Spectrum of heart disease and risk factors in a black urban population in South Africa (the Heart of Soweto Study): a cohort study. *Lancet*. 2008;371:915–922. doi: 10.1016/S0140-6736(08)60417-1
- 40. Prattipati S, Sakita FM, Tarimo TG, Kweka GL, Mlangi JJ, Maro AV, Coaxum LA, Galson SW, Limkakeng AT, Rugakingira A, et al. Prevalence and correlates of ischemic ECG findings among adults with and without HIV in Tanzania. *Glob Heart*. 2022;17:38. doi: 10.5334/gh.1127
- 41. Fung G, Luo H, Qiu Y, Yang D, McManus B. Myocarditis. *Circ Res*. 2016;118:496–514. doi: 10.1161/CIRCRESAHA.115.306573
- 42. Shaboodien G, Maske C, Wainwright H, Smuts H, Ntsekhe M, Commerford PJ, Badri M, Mayosi BM. Prevalence of myocarditis and cardiotropic virus infection in Africans with HIV-associated cardiomyopathy, idiopathic dilated cardiomyopathy and heart transplant recipients: a pilot study: cardiovascular topic. *Cardiovasc J Afr*. 2013;24:218–223. doi: 10.5830/CVJA-2013-039
- 43. Fourie CMT, Schutte AE, Smith W, Kruger A, van Rooyen JM. Endothelial activation and cardiometabolic profiles of treated and nevertreated HIV infected Africans. *Atherosclerosis*. 2015;240:154–160. doi: 10.1016/j.atherosclerosis.2015.03.015
- 44. Mosepele M, Hemphill LC, Moloi W, Moyo S, Nkele I, Makhema J, Bennett K, Triant VA, Lockman S. Pre-clinical carotid atherosclerosis and sCD163 among virally suppressed HIV patients in Botswana compared with uninfected controls. *PLoS One*. 2017;12:e0179994. doi: 10.1371/journal.pone.0179994
- 45. Alencherry B, Erem G, Mirembe G, Ssinabulya I, Yun CH, Hung CL, Siedner MJ, Bittencourt M, Kityo C, McComsey GA, et al. Coronary artery calcium, HIV and inflammation in Uganda compared with the USA. *Open Heart*. 2019;6:e001046. doi: 10.1136/openhrt-2019-001046
- 46. Nonterah EA, Boua PR, Klipstein-Grobusch K, Asiki G, Micklesfield LK, Agongo G, Ali SA, Mashinya F, Sorgho H, Nakanabo-Diallo S, et al; as members and collaborators of AWI-Gen and the H3Africa Consortium. Classical cardiovascular risk factors and HIV are associated with carotid intima-media thickness in adults from sub-Saharan Africa: findings from H3Africa AWI-Gen study. *J Am Heart Assoc*. 2019;8:e011506. doi: 10.1161/JAHA.118.011506
- 47. Shuldiner SR, Wong LY, Peterson TE, Wolfson J, Jermy S, Saad H, Lumbamba MAJ, Singh A, Shey M, Meintjes G, et al. Myocardial fibrosis among antiretroviral therapy-treated persons with human immunodeficiency virus in South Africa. *Open Forum Infect Dis*. 2021;8:ofaa600. doi: 10.1093/ofid/ofaa600
- 48. Robbertse PPS, Doubell AF, Lombard CJ, Talle MA, Herbst PG. Evolution of myocardial oedema and fibrosis in HIV infected persons after the initiation of antiretroviral therapy: a prospective cardiovascular magnetic resonance study. *J Cardiovasc Magn Reson*. 2022;24:72. doi: 10.1186/s12968-022-00901-0
- 49. Peterson TE, Shey M, Masina N, Wong LY, Shuldiner SR, Wolfson J, Jermy S, Saad H, Lumbamba MAJ, Singh A, et al. Myocardial extracellular volume fraction is positively associated with activated monocyte subsets among cART-treated persons living with HIV in South Africa. *Int J Cardiol*. 2023;392:131332. doi: 10.1016/j.ijcard.2023.131332
- 50. Longenecker CT, Bogorodskaya M, Margevicius S, Nazzinda R, Bittencourt MS, Erem G, Nalukwago S, Huaman MA, Ghoshhajra BB, Siedner MJ, et al. Sex modifies the association between HIV and coronary artery disease among older adults in Uganda. *J Int AIDS Soc*. 2022;25:e25868. doi: 10.1002/jia2.25868
- 51. Lawal IO, Ankrah AO, Popoola GO, Lengana T, Sathekge MM. Arterial inflammation in young patients with human immunodeficiency virus infection: a cross-sectional study using F-18 FDG PET/CT. *J Nucl Cardiol*. 2019;26:1258–1265. doi: 10.1007/s12350-018-1207-x
- 52. Ssinabulya I, Kayima J, Longenecker C, Luwedde M, Semitala F, Kambugu A, Ameda F, Bugeza S, McComsey G, Freers J, et al. Subclinical atherosclerosis among HIV-infected adults attending HIV/AIDS care at two large ambulatory HIV clinics in Uganda. *PLoS One*. 2014;9:e89537. doi: 10.1371/journal.pone.0089537
- 53. Siedner MJ, Bibangambah P, Kim JH, Lankowski A, Chang JL, Yang IT, Kwon DS, North CM, Triant VA, Longenecker C, et al. Treated HIV infection and progression of carotid atherosclerosis in rural Uganda: a prospective observational cohort study. *J Am Heart Assoc*. 2021;10:e019994. doi: 10.1161/JAHA.120.019994
- 54. Dirajlal-Fargo S, Sattar A, Yu J, Albar Z, Chaves FC, Riedl K, Kityo C, Bowman E, McComsey GA, Funderburg N. Lipidome association with vascular disease and inflammation in HIV+ Ugandan children. *AIDS*. 2021;35:1615–1623. doi: 10.1097/QAD.0000000000002923
- 55. Fourie C, van Rooyen J, Pieters M, Conradie K, Hoekstra T, Schutte A. Is HIV-1 infection associated with endothelial dysfunction in a population of African ancestry in South Africa? *Cardiovasc J Afr*. 2011;22:134–140. doi: 10.5830/cvja-2010-056
- 56. Huaman MA, Feria MG, Kityo C, Nalukwago S, Nazzinda R, Zidar DA, Zanni MV, Siedner MJ, Grinspoon SK, Longenecker CT. A sex-stratified analysis of monocyte phenotypes associated with HIV infection in Uganda. *Viruses*. 2021;13:2135. doi: 10.3390/v13112135
- 57. Shakil SS, Temu TM, Kityo C, Nazzinda R, Erem G, Kentoffio K, Bittencourt M, Ntusi NAB, Zanni MV, Longenecker CT. Sex modulates the association between inflammation and coronary atherosclerosis among older Ugandan adults with and without HIV. *AIDS*. 2023;37:579–586. doi: 10.1097/QAD.0000000000003451
- 58. Grody WW, Cheng L, Lewis W. Infection of the heart by the human immunodeficiency virus. *Am J Cardiol*. 1990;66:203–206. doi: 10.1016/0002-9149(90)90589-s
- 59. Lipshultz SE, Fox CH, Perez-Atayde AR, Sanders SP, Colan SD, McIntosh K, Winter HS. Identification of human immunodeficiency virus-1 RNA and DNA in the heart of a child with cardiovascular abnormalities and congenital acquired immune deficiency syndrome. *Am J Cardiol*. 1990;66:246–250. doi: 10.1016/0002-9149(90)90603-x
- 60. Herskowitz A, Wu TC, Willoughby SB, Vlahov D, Ansari AA, Beschorner WE, Baughman KL. Myocarditis and cardiotropic viral infection associated with severe left ventricular dysfunction in late-stage infection with human immunodeficiency virus. *J Am Coll Cardiol*. 1994;24:1025–1032. doi: 10.1016/0735-1097(94)90865-6
- 61. Twu C, Liu NQ, Popik W, Bukrinsky M, Sayre J, Roberts J, Rania S, Bramhandam V, Roos KP, MacLellan WR, et al. Cardiomyocytes undergo apoptosis in human immunodeficiency virus cardiomyopathy through mitochondrion- and death receptor-controlled pathways. *Proc Natl Acad Sci USA*. 2002;99:14386–14391. doi: 10.1073/pnas.212327899
- 62. Rebolledo MA, Krogstad P, Chen F, Shannon KM, Klitzner TS. Infection of human fetal cardiac myocytes by a human immunodeficiency virus-1– derived vector. *Circ Res*. 1998;83:738–742. doi: 10.1161/01.res.83.7.738
- 63. Fiala M, Popik W, Qiao JH, Lossinsky AS, Alce T, Tran K, Yang W, Roos KP, Arthos J. HIV-1 induces cardiomyopathyby cardiomyocyte invasion and gp120, Tat, and cytokine apoptotic signaling. *Cardiovasc Toxicol*. 2004;4:97– 107. doi: 10.1385/ct:4:2:097
- 64. Shannon RP, Mathier MA, Manleod S. Macrophages not cardiomyocytes are the reservoir for lentivirus in SIV cardiomyopathy. *Circulation*. 1999;100(suppl I):1–382.
- 65. Yoon V, Fridkis-Hareli M, Munisamy S, Lee J, Anastasiades D, Stevceva L. The GP120 molecule of HIV-1 and its interaction with T cells. *Curr Med Chem*. 2010;17:741–749. doi: 10.2174/092986710790514499
- 66. Ahmad N, Venkatesan S. Nef protein of HIV-1 is a transcriptional repressor of HIV-1 LTR. *Science*. 1988;241:1481–1485. doi: 10.1126/science.3262235
- 67. Gupta MK, Kaminski R, Mullen B, Gordon J, Burdo TH, Cheung JY, Feldman AM, Madesh M, Khalili K. HIV-1 Nef-induced cardiotoxicity through dysregulation of autophagy. *Sci Rep*. 2017;7:8572. doi: 10.1038/s41598-017-08736-x
- 68. Raymond AD, Campbell-Sims TC, Khan M, Lang M, Huang MB, Bond VC, Powell MD. HIV type 1 Nef is released from infected cells in CD45+ microvesicles and is present in the plasma of HIV-infected individuals. *AIDS Res Hum Retroviruses*. 2011;27:167–178. doi: 10.1089/aid.2009.0170
- 69. Fang Q, Kan H, Lewis W, Chen F, Sharma P, Finkel MS. Dilated cardiomyopathy in transgenic mice expressing HIV Tat. *Cardiovasc Toxicol*. 2009;9:39– 45. doi: 10.1007/s12012-009-9035-5
- 70. Barbaro G, Di Lorenzo G, Grisorio B, Barbarini G. Cardiac involvement in the acquired immunodeficiency syndrome: a multicenter clinicalpathological study. *AIDS Res Hum Retroviruses*. 1998;14:1071–1077. doi: 10.1089/aid.1998.14.1071
- 71. Low A, Gavriilidis G, Larke N, B-Lajoie MR, Drouin O, Stover J, Muhe L, Easterbrook P. Incidence of opportunistic infections and the impact of antiretroviral therapy among HIV-infected adults in low- and middle-income countries: a systematic review and meta-analysis. *Clin Infect Dis*. 2016;62:1595–1603. doi: 10.1093/cid/ciw125
- 72. Tschöpe C, Ammirati E, Bozkurt B, Caforio ALP, Cooper LT, Felix SB, Hare JM, Heidecker B, Heymans S, Hübner N, et al. Myocarditis and inflammatory cardiomyopathy: current evidence and future directions. *Nat Rev Cardiol*. 2021;18:169–193. doi: 10.1038/s41569-020-00435-x
- 73. Kedzierska K, Crowe SM. Cytokines and HIV-1: interactions and clinical implications. *Antiviral Chem Chemother*. 2001;12:133–150. doi: 10.1177/095632020101200301
- 74. Monsuez JJ, Escaut L, Teicher E, Charniot JC, Vittecoq D. Cytokines in HIV-associated cardiomyopathy. *Int J Cardiol*. 2007;120:150–157. doi: 10.1016/j.ijcard.2006.11.143
- 75. Barbaro G, Di Lorenzo G, Soldini M, Giancaspro G, Grisorio B, Pellicelli A, Barbarini G. Intensity of myocardial expression of inducible nitric oxide synthase influences the clinical course of human immunodeficiency virusassociated cardiomyopathy. *Circulation*. 1999;100:933–939. doi: 10.1161/01.cir.100.9.933
- 76. Kan H, Xie Z, Finkel MS. HIV gp120 enhances NO production by cardiac myocytes through p38 MAP kinase-mediated NF-κB activation. *Am J Physiol Heart Circ Physiol*. 2000;279:H3138–H3143. doi: 10.1152/ajpheart.2000.279.6.h3138
- 77. Kelly KM, Tocchetti CG, Lyashkov A, Tarwater PM, Bedja D, Graham DR, Beck SE, Metcalf Pate KA, Queen SE, Adams RJ, et al. CCR5 inhibition prevents cardiac dysfunction in the SIV/macaque model of HIV. *J Am Heart Assoc*. 2014;3:e000874. doi: 10.1161/JAHA.114.000874
- 78. Luetkens JA, Doerner J, Schwarze-Zander C, Wasmuth JC, Boesecke C, Sprinkart AM, Schmeel FC, Homsi R, Gieseke J, Schild HH, et al. Cardiac magnetic resonance reveals signs of subclinical myocardial inflammation in asymptomatic HIV-Infected patients. *Circ Cardiovasc Imaging*. 2016;9:e004091. doi: 10.1161/CIRCIMAGING.115.004091
- 79. Yan C, Li R, Guo X, Yu H, Li W, Li W, Ren M, Yang M, Li H. Cardiac involvement in human immunodeficiency virus infected patients: an observational cardiac magnetic resonance study. *Front Cardiovasc Med*. 2021;8:756162. doi: 10.3389/FCVM.2021.756162
- 80. Menacho K, Seraphim A, Ramirez S, Falcon L, Bhuva A, Alave J, Banda C, Mejia F, Salazar D, Putri A, et al. Myocardial inflammation and edema in people living with human immunodeficiency virus. *JACC Cardiovasc Imaging*. 2020;13:1278–1280. doi: 10.1016/j.jcmg.2019.12.005
- 81. Zanni MV, Awadalla M, Toribio M, Robinson J, Stone LA, Cagliero D, Rokicki A, Mulligan CP, Ho JE, Neilan AM, et al. Immune correlates of diffuse myocardial fibrosis and diastolic dysfunction among aging women with human immunodeficiency virus. *J Infect Dis*. 2020;221:1315–1320. doi: 10.1093/infdis/jiz184
- 82. Wu KC, Haberlen SA, Plankey MW, Palella FJ, Piggott DA, Kirk GD, Margolick JB, Post WS. Human immunodeficiency viral infection and differences in interstitial ventricular fibrosis and left atrial size. *Eur Heart J Cardiovasc Imaging*. 2021;22:888–895. doi: 10.1093/ehjci/jeab037
- 83. Ntusi N, O'Dwyer E, Dorrell L, Wainwright E, Piechnik S, Clutton G, Hancock G, Ferreira V, Cox P, Badri M, et al. HIV-1-related cardiovascular disease is associated with chronic inflammation, frequent pericardial effusions, and probable myocardial edema. *Circ Cardiovasc Imaging*. 2016;9:e004430. doi: 10.1161/CIRCIMAGING.115.004430
- 84. Alam SR, Vinayak S, Shah A, Doolub G, Kimeu R, Horn KP, Bowen SR, Jeilan M, Lee KK, Gachoka S, et al. Assessment of cardiac, vascular, and pulmonary pathobiology in vivo during acute COVID-19. *J Am Heart Assoc*. 2022;11:e026399. doi: 10.1161/JAHA.122.026399
- 85. Alam SR, Shah ASV, Ombati KO, Nganga E, Gitau Makhdomi K, Chung MH, Vinayak S. CardiOvaScular Mechanisms In Covid-19: methodology of a prospective observational multimodality imaging study (COSMIC-19 study). *BMC Cardiovasc Disord*. 2021;21:234. doi: 10.1186/s12872-021-02027-0
- 86. Abplanalp WT, Tucker N, Dimmeler S. Single-cell technologies to decipher cardiovascular diseases. *Eur Heart J*. 2022;43:4536–4547. doi: 10.1093/eurheartj/ehac095
- 87. Koenig AL, Shchukina I, Amrute J, Andhey PS, Zaitsev K, Lai L, Bajpai G, Bredemeyer A, Smith G, Jones C, et al. Single-cell transcriptomics reveals cell-type-specific diversification in human heart failure. *Nat Cardiovasc Res*. 2022;1:263–280. doi: 10.1038/s44161-022-00028-6
- 88. Tombor LS, John D, Glaser SF, Luxán G, Forte E, Furtado M, Rosenthal N, Baumgarten N, Schulz MH, Wittig J, et al. Single cell sequencing reveals endothelial plasticity with transient mesenchymal activation after myocardial infarction. *Nat Commun*. 2021;12:681. doi: 10.1038/s41467-021-20905-1
- 89. Vallejo J, Cochain C, Zernecke A, Ley K. Heterogeneity of immune cells in human atherosclerosis revealed by scRNA-Seq. *Cardiovasc Res*. 2021;117:2537–2543. doi: 10.1093/cvr/cvab260
- 90. Sun J, Singh P, Shami A, Kluza E, Pan M, Djordjevic D, Michaelsen NB, Kennbäck C, van der Wel NN, Orho-Melander M, et al. Spatial transcriptional mapping reveals site-specific pathways underlying human atherosclerotic plaque rupture. *J Am Coll Cardiol*. 2023;81:2213–2227. doi: 10.1016/j.jacc.2023.04.008
- 91. Mantri M, Hinchman MM, McKellar DW, Wang MFZ, Cross ST, Parker JSL, De Vlaminck I. Spatiotemporal transcriptomics reveals pathogenesis of viral myocarditis. *Nat Cardiovasc Res*. 2022;1:946–960. doi: 10.1038/s44161-022-00138-1
- 92. Theofilatos K, Stojkovic S, Hasman M, van der Laan SW, Baig F, Barallobre-Barreiro J, Schmidt LE, Yin S, Yin X, Burnap S, et al. Proteomic atlas of atherosclerosis: the contribution of proteoglycans to sex differences, plaque phenotypes, and outcomes. *Circ Res*. 2023;133:542–558. doi: 10.1161/CIRCRESAHA.123.322590
- 93. Bhatia HS, Brunner AD, Öztürk F, Kapoor S, Rong Z, Mai H, Thielert M, Ali M, Al-Maskari R, Paetzold JC, et al. Spatial proteomics in three-dimensional intact specimens. *Cell*. 2022;185:5040–5058.e19. doi: 10.1016/j.cell.2022.11.021
- 94. Holla P, Dizon B, Ambegaonkar AA, Rogel N, Goldschmidt E, Boddapati AK, Sohn H, Sturdevant D, Austin JW, Kardava L, et al. Shared transcriptional profiles of atypical B cells suggest common drivers of expansion and function in malaria, HIV, and autoimmunity. *Sci Adv*. 2021;7:eabg8384. doi: 10.1126/sciadv.abg8384
- 95. Zhang X, Qazi AA, Deshmukh S, Lobato Ventura R, Mukim A, Beliakova-Bethell N. Single-cell RNA sequencing reveals common and unique gene expression profiles in primary CD4+ T cells latently infected with HIV under different conditions. *Front Cell Infect Microbiol*. 2023;13:1286168. doi: 10.3389/fcimb.2023.1286168
- 96. Cham LB, Gunst JD, Schleimann MH, Frattari GS, Rosas-Umbert M, Vibholm LK, van der Sluis RM, Jakobsen MR, Olesen R, Lin L, et al. Single cell analysis reveals a subset of cytotoxic-like plasmacytoid dendritic cells in people with HIV-1. *iScience*. 2023;26:107628. doi: 10.1016/j.isci.2023.107628
- 97. Kong P, Christia P, Frangogiannis NG. The pathogenesis of cardiac fibrosis. *Cell Mol Life Sci*. 2014;71:549–574. doi: 10.1007/s00018-013-1349-6
- 98. Tseng ZH, Secemsky EA, Dowdy D, Vittinghoff E, Moyers B, Wong JK, Havlir DV, Hsue PY. Sudden cardiac death in patients with human immunodeficiency virus infection. *J Am Coll Cardiol*. 2012;59:1891–1896. doi: 10.1016/j.jacc.2012.02.024
- 99. Thomas TP, Grisanti LA. The dynamic interplay between cardiac inflammation and fibrosis. *Front Physiol*. 2020;11:529075. doi: 10.3389/fphys.2020.529075
- 100. Teer E, Dominick L, Mukonowenzou NC, Essop MF. HIV-related myocardial fibrosis: inflammatory hypothesis and crucial role of immune cells dysregulation. *Cells*. 2022;11:2825. doi: 10.3390/cells11182825
- 101. Cai CW, Sereti I. Residual immune dysfunction under antiretroviral therapy. *Semin Immunol*. 2021;51:101471. doi: 10.1016/j.smim.2021.101471
- 102. Hulsmans M, Sam F, Nahrendorf M. Monocyte and macrophage contributions to cardiac remodeling. *J Mol Cell Cardiol*. 2016;93:149–155. doi: 10.1016/j.yjmcc.2015.11.015
- 103. Yu Y, Schürpf T, Springer TA. How natalizumab binds and antagonizes α4 integrins. *J Biol Chem*. 2013;288:32314–32325. doi: 10.1074/jbc.M113.501668
- 104. Walker JA, Beck GA, Campbell JH, Miller AD, Burdo TH, Williams KC. Anti-α4 integrin antibody blocks monocyte/macrophage traffic to the heart and decreases cardiac pathology in a SIV infection model of AIDS. *J Am Heart Assoc*. 2015;4:e001932. doi: 10.1161/JAHA.115.001932
- 105. Burdo TH, Walker J, Williams KC. Macrophage polarization in AIDS: dynamic interface between anti-viral and anti-inflammatory macrophages during acute and chronic infection. *J Clin Cell Immunol*. 2015;6:333.
- 106. Meng X, Nikolic-Paterson DJ, Lan HY. TGF-β: the master regulator of fibrosis. *Nat Rev Nephrol*. 2016;12:325–338. doi: 10.1038/nrneph.2016.48

HIV AND CARDIOVASCULAR<br>Disease Review Series

- 107. Wiercińska-Drapalo A, Flisiak R, Jaroszewicz J, Prokopowicz D. Increased plasma transforming growth factor-beta1 is associated with disease progression in HIV-1-infected patients. *Viral Immunol*. 2004;17:109–113. doi: 10.1089/088282404322875502
- 108. Musa F, Shaviya N, Mambo F, Abonyo C, Barasa E, Wafula P, Sowayi G, Barasa M, Were T. Cytokine profiles in highly active antiretroviral treatment non-adherent, adherent and naive HIV-1 infected patients in Western Kenya. *Afr Health Sci*. 2021;21:1584–1592. doi: 10.4314/ahs.v21i4.12
- 109. Jiang Y, Chai L, Wang H, Shen X, Fasae MB, Jiao J, Yu Y, Ju J, Liu B, Bai Y. HIV tat protein induces myocardial fibrosis through TGF-β1-CTGF signaling cascade: a potential mechanism of HIV infection-related cardiac manifestations. *Cardiovasc Toxicol*. 2021;21:965–972. doi: 10.1007/s12012-021-09687-6
- 110. Laurence J, Elhadad S, Robison T, Terry H, Varshney R, Woolington S, Ghafoory S, Choi ME, Ahamed J. HIV protease inhibitor-induced cardiac dysfunction and fibrosis is mediated by platelet-derived TGF-β1 and can be suppressed by exogenous carbon monoxide. *PLoS One*. 2017;12:e0187185. doi: 10.1371/journal.pone.0187185
- 111. Zaborska B, Sikora-Frąc M, Smarż K, Pilichowska-Paszkiet E, Budaj A, Sitkiewicz D, Sygitowicz G. The role of galectin-3 in heart failure—the diagnostic, prognostic and therapeutic potential—where do we stand? *Int J Mol Sci*. 2023;24:13111. doi: 10.3390/ijms241713111
- 112. Gao Z, Liu Z, Wang R, Zheng Y, Li H, Yang L. Galectin-3 is a potential mediator for atherosclerosis. *J Immunol Res*. 2020;2020:5284728. doi: 10.1155/2020/5284728
- 113. Lee KN, Kim DY, Boo KY, Kim YG, Roh SY, Baek YS, Kim DH, Lee DI, Shim J, Choi JI, et al. Therapeutic implications of galectin-3 in patients with atrial fibrillation. *Sci Rep*. 2022;12:784. doi: 10.1038/s41598-022-04894-9
- 114. Luo J, Wang S, Liu X, Zheng Q, Wang Z, Huang Y, Shi J. Galectin-3 promotes calcification of human aortic valve interstitial cells via the NF-kappa B signaling pathway. *Cardiovasc Diagn Ther*. 2022;12:196–207. doi: 10.21037/cdt-21-506
- 115. Sciacchitano S, Lavra L, Morgante A, Ulivieri A, Magi F, De Francesco GP, Bellotti C, Salehi LB, Ricci A. Galectin-3: one molecule for an alphabet of diseases, from A to Z. *Int J Mol Sci*. 2018;19:379. doi: 10.3390/ijms19020379
- 116. Shah RV, Chen-Tournoux AA, Picard MH, van Kimmenade RRJ, Januzzi JL. Serum levels of the interleukin-1 receptor family member ST2, cardiac structure and function, and long-term mortality in patients with acute dyspnea. *Circ Heart Fail*. 2009;2:311–319. doi: 10.1161/CIRCHEARTFAILURE.108.833707
- 117. Fogel S, Guittaut M, Legrand A, Monsigny M, Hebert E. The Tat protein of HIV-1 induces galectin-3 expression. *Glycobiology*. 1999;9:383–387. doi: 10.1093/glycob/9.4.383
- 118. Martínez-Martínez E, Brugnolaro C, Ibarrola J, Ravassa Buonafine M, López B, Fernández-Celis A, Querejeta R, Santamaria E, Fernández-Irigoyen J, et al. CT-1 (cardiotrophin-1)-Gal-3 (galectin-3) axis in cardiac fibrosis and inflammation. *Hypertension*. 2019;73:602– 611. doi: 10.1161/HYPERTENSIONAHA.118.11874
- 119. Pascual-Figal DA, Januzzi JL. The biology of ST2: the International ST2 Consensus Panel. *Am J Cardiol*. 2015;115:3B–7B. doi: 10.1016/j.amjcard.2015.01.034
- 120. Sanada S, Hakuno D, Higgins LJ, Schreiter E, McKenzie A, Lee RT. IL-33 and ST2 comprise a critical biomechanically induced and cardioprotective signaling system. *J Clin Invest*. 2007;117:1538–1549. doi: 10.1172/JCI30634
- 121. Wang Z, Pan X, Xu H, Wu Y, Jia X, Fang Y, Lu Y, Xu Y, Zhang J, Su Y. Serum soluble ST2 is a valuable prognostic biomarker in patients with acute heart failure. *Front Cardiovasc Med*. 2022;9:812654. doi: 10.3389/fcvm.2022.812654
- 122. Secemsky EA, Scherzer R, Nitta E, Wu AHB, Lange DC, Deeks SG, Martin JN, Snider J, Ganz P, Hsue PY. Novel biomarkers of cardiac stress, cardiovascular dysfunction, and outcomes in HIV-Infected individuals. *JACC Heart Failure*. 2015;3:591–599. doi: 10.1016/j.jchf.2015.03.007
- 123. Robinson JA, Toribio M, Quinaglia T, Awadalla M, Talathi R, Durbin CG, Alhallak I, Alagpulinsa DA, Fourman LT, Suero-Abreu GA, et al. Plasma osteopontin relates to myocardial fibrosis and steatosis and to immune activation among women with HIV. *AIDS*. 2023;37:305–310. doi: 10.1097/QAD.0000000000003417
- 124. Soehnlein O, Libby P. Targeting inflammation in atherosclerosis from experimental insights to the clinic. *Nat Rev Drug Discov*. 2021;20:589–610. doi: 10.1038/s41573-021-00198-1
- 125. Welsh P, Grassia G, Botha S, Sattar N, Maffia P. Targeting inflammation to reduce cardiovascular disease risk: a realistic clinical prospect? *Br J Pharmacol*. 2017;174:3898–3913. doi: 10.1111/bph.13818
- 126. So-Armah K, Benjamin LA, Bloomfield GS, Feinstein MJ, Hsue P, Njuguna B, Freiberg MS. HIV and cardiovascular disease. *Lancet HIV*. 2020;7:e279– e293. doi: 10.1016/S2352-3018(20)30036-9
- 127. Tawakol A, Migrino RQ, Bashian GG, Bedri S, Vermylen D, Cury RC, Yates D, LaMuraglia GM, Furie K, Houser S, et al. In vivo 18F-fluorodeoxyglucose positron emission tomography imaging provides a noninvasive measure of carotid plaque inflammation in patients. *J Am Coll Cardiol*. 2006;48:1818– 1824. doi: 10.1016/j.jacc.2006.05.076
- 128. Figueroa AL, Abdelbaky A, Truong QA, Corsini E, MacNabb MH, Lavender ZR, Lawler MA, Grinspoon SK, Brady TJ, Nasir K, et al. Measurement of arterial activity on routine FDG PET/CT images improves prediction of risk of future CV events. *JACC Cardiovasc Imaging*. 2013;6:1250–1259. doi: 10.1016/j.jcmg.2013.08.006
- 129. Hetterich H, Rominger A, Walter L, Habs M, Volpers S, Hacker M, Reiser MF, Bartenstein P, Saam T. Natural history of atherosclerotic disease progression as assessed by (18)F-FDG PET/CT. *Int J Cardiovasc Imaging*. 2016;32:49–59. doi: 10.1007/s10554-015-0660-8
- 130. Longenecker CT, Sullivan CE, Morrison J, Hileman CO, Zidar DA, Gilkeson R, O'Donnell J, McComsey GA. The effects of HIV and smoking on aortic and splenic inflammation. *AIDS*. 2018;32:89–94. doi: 10.1097/QAD.0000000000001682
- 131. Knudsen A, Hag AMF, Loft A, von Benzon E, Keller SH, Møller HJ, Lebech AM, Ripa RS, Kjær A. HIV infection and arterial inflammation assessed by 18F-fluorodeoxyglucose (FDG) positron emission tomography (PET): a prospective cross-sectional study. *J Nucl Cardiol*. 2015;22:372– 380. doi: 10.1007/s12350-014-0032-0
- 132. Yarasheski KE, Laciny E, Overton E, Reeds DN, Harrod M, Baldwin S, Dávila-Román VG. 18FDG PET-CT imaging detects arterial inflammation and early atherosclerosis in HIV-infected adults with cardiovascular disease risk factors. *J Inflamm*. 2012;9:26. doi: 10.1186/1476-9255-9-26
- 133. Lo J, Lu MT, Ihenachor EJ, Wei J, Looby SE, Fitch KV, Oh J, Zimmerman CO, Hwang J, Abbara S, et al. Effects of statin therapy on coronary artery plaque volume and high-risk plaque morphology in HIV-infected patients with subclinical atherosclerosis: a randomised, double-blind, placebo-controlled trial. *Lancet HIV*. 2015;2:e52–e63. doi: 10.1016/S2352-3018(14)00032-0
- 134. Enriquez R, Ssekubugu R, Ndyanabo A, Marrone G, Gigante B, Chang LW, Reynolds SJ, Nalugoda F, Ekstrom AM, Sewankambo NK, et al. Prevalence of cardiovascular risk factors by HIV status in a population-based cohort in South Central Uganda: a cross-sectional survey. *J Int AIDS Soc*. 2022;25:e25901. doi: 10.1002/jia2.25901
- 135. Manne-Goehler J, Montana L, Gómez-Olivé FX, Rohr J, Harling G, Wagner RG, Wade A, Kabudula CW, Geldsetzer P, Kahn K, et al. The ART advantage: healthcare utilization for diabetes and hypertension in rural South Africa. *J Acquir Immune Defic Syndr*. 2017;75:561–567. doi: 10.1097/QAI.0000000000001445
- 136. Post WS, Budoff M, Kingsley L, Palella FJ, Witt MD, Li X, George RT, Brown TT, Jacobson LP. Associations between HIV infection and subclinical coronary atherosclerosis. *Ann Intern Med*. 2014;160:458–467. doi: 10.7326/M13-1754
- 137. Boldeanu I, Sadouni M, Mansour S, Baril JG, Trottier B, Soulez G, S Chin A, Leipsic J, Tremblay C, Durand M, et al; Canadian HIV and Aging Cohort Study Group. Prevalence and characterization of subclinical coronary atherosclerotic plaque with CT among individuals with HIV: results from the Canadian HIV and aging cohort study. *Radiology*. 2021;299:571–580. doi: 10.1148/radiol.2021203297
- 138. Tarr PE, Ledergerber B, Calmy A, Doco-Lecompte T, Marzel A, Weber R, Kaufmann PA, Nkoulou R, Buechel RR, Kovari H; Swiss HIV Cohort Study. Subclinical coronary artery disease in Swiss HIV-positive and HIV-negative persons. *Eur Heart J*. 2018;39:2147–2154. doi: 10.1093/eurheartj/ehy163
- 139. Lai H, Moore R, Celentano DD, Gerstenblith G, Treisman G, Keruly JC, Kickler T, Li J, Chen S, Lai S, et al. HIV infection itself may not be associated with subclinical coronary artery disease among African Americans without cardiovascular symptoms. *J Am Heart Assoc*. 2015;5:e002529. doi: 10.1161/JAHA.115.002529
- 140. Li P, Zhang Y, Gao J, Zhao Q. Observation on vulnerable coronary plaques in HIV-infected patients based on coronary CTA. *Chin J Med Imag Technol*. 2020;36:391–395. doi: 10.13929/j.issn.1003-3289.2020.03.017
- 141. Feinstein MJ, Kim JH, Bibangambah P, Sentongo R, Martin JN, Tsai AC, Bangsberg DR, Hemphill L, Triant VA, Boum Y, et al. Ideal cardiovascular health and carotid atherosclerosis in a mixed cohort of HIV-infected and uninfected Ugandans. *AIDS Res Hum Retroviruses*. 2017;33:49–56. doi: 10.1089/AID.2016.0104
- 142. Neuhaus J, Jacobs DR, Baker JV, Calmy A, Duprez D, La Rosa A, Kuller LH, Pett SL, Ristola M, Ross MJ, et al. Markers of inflammation, coagulation,

**HIV AND CARDIOVASCULAR**  IIV AND CARDIOVASCULAR<br>Disease review series **DISEASE REVIEW SERIES**

and renal function are elevated in adults with HIV infection. *J Infect Dis*. 2010;201:1788–1795. doi: 10.1086/652749

- 143. Perkins MV, Joseph SB, Dittmer DP, Mackman N. Cardiovascular disease and thrombosis in HIV infection. *Arterioscler Thromb Vasc Biol*. 2023;43:175–191. doi: 10.1161/ATVBAHA.122.318232
- 144. Kuller LH, Tracy R, Belloso W, Wit SD, Drummond F, Lane HC, Ledergerber B, Lundgren J, Neuhaus J, Nixon D, et al; for the INSIGHT SMART Study Group. Inflammatory and coagulation biomarkers and mortality in patients with HIV infection. *PLoS Med*. 2008;5:e203. doi: 10.1371/journal.pmed.0050203
- 145. Nordell AD, McKenna M, Borges AH, Duprez D, Neuhaus J, Neaton JD; INSIGHT SMART, ESPRIT Study Groups. Severity of cardiovascular disease outcomes among patients with HIV is related to markers of inflammation and coagulation. *J Am Heart Assoc*. 2014;3:e000844. doi: 10.1161/JAHA.114.000844
- 146. Freeman ML, Hossain MB, Burrowes SAB, Jeudy J, Bui R, Moisi D, Mitchell SE, Khambaty M, Weiss RG, Lederman MM, et al. Association of soluble markers of inflammation with peri-coronary artery inflammation in people with and without HIV infection and without cardiovascular disease. *Open Forum Infect Dis*. 2023;10:ofad328. doi: 10.1093/ofid/ofad328
- 147. Hsue PY, Waters DD. HIV infection and coronary heart disease: mechanisms and management. *Nat Rev Cardiol*. 2019;16:745–759. doi: 10.1038/s41569-019-0219-9
- 148. van der Heijden WA, Van de Wijer L, Keramati F, Trypsteen W, Rutsaert S, Horst RT, Jaeger M, Koenen HJ, Stunnenberg HG, Joosten I, et al. Chronic HIV infection induces transcriptional and functional reprogramming of innate immune cells. *JCI Insight*. 2021;6:e145928. doi: 10.1172/jci.insight.145928
- 149. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, Fonseca F, Nicolau J, Koenig W, Anker SD, et al; CANTOS Trial Group. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med*. 2017;377:1119–1131. doi: 10.1056/NEJMoa1707914
- 150. Ridker PM, Devalaraja M, Baeres FMM, Engelmann MDM, Hovingh GK, Ivkovic M, Lo L, Kling D, Pergola P, Raj D, et al; RESCUE Investigators. IL-6 inhibition with ziltivekimab in patients at high atherosclerotic risk (RESCUE): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet*. 2021;397:2060–2069. doi: 10.1016/S0140-6736(21)00520-1
- 151. Santinelli L, De Girolamo G, Borrazzo C, Vassalini P, Pinacchio C, Cavallari EN, Statzu M, Frasca F, Scordio M, Bitossi C, et al. Alteration of type I interferon response is associated with subclinical atherosclerosis in virologically suppressed HIV-1-infected male patients. *J Med Virol*. 2021;93:4930–4938. doi: 10.1002/jmv.27028
- 152. Wanjalla CN, Temu TM, Mashayekhi M, Warren CM, Shepherd BE, Gangula R, Fuseini H, Bailin S, Gabriel CL, Gangula P, et al. Interleukin-17A is associated with flow-mediated dilation and interleukin-4 with carotid plaque in persons with HIV. *AIDS*. 2022;36:963–973. doi: 10.1097/QAD.0000000000003196
- 153. Zhang H, Yang K, Chen F, Liu Q, Ni J, Cao W, Hua Y, He F, Liu Z, Li L, et al. Role of the CCL2-CCR2 axis in cardiovascular disease: pathogenesis and clinical implications. *Front Immunol*. 2022;13:975367. doi: 10.3389/fimmu.2022.975367
- 154. Mikolajczyk TP, Szczepaniak P, Vidler F, Maffia P, Graham GJ, Guzik TJ. Role of inflammatory chemokines in hypertension. *Pharmacol Ther*. 2021;223:107799. doi: 10.1016/j.pharmthera.2020.107799
- 155. Floris-Moore M, Fayad ZA, Berman JW, Mani V, Schoenbaum EE, Klein RS, Weinshelbaum KB, Fuster V, Howard AA, Lo Y, et al. Association of HIV viral load with monocyte chemoattractant protein-1 and atherosclerosis burden measured by magnetic resonance imaging. *AIDS*. 2009;23:941–949. doi: 10.1097/QAD.0b013e328329c76b
- 156. Burdo TH, Lo J, Abbara S, Wei J, DeLelys ME, Preffer F, Rosenberg ES, Williams KC, Grinspoon S. Soluble CD163, a novel marker of activated macrophages, is elevated and associated with noncalcified coronary plaque in HIV-infected patients. *J Infect Dis*. 2011;204:1227–1236. doi: 10.1093/infdis/jir520
- 157. Fitch KV, Srinivasa S, Abbara S, Burdo TH, Williams KC, Eneh P, Lo J, Grinspoon SK. Noncalcified coronary atherosclerotic plaque and immune activation in HIV-infected women. *J Infect Dis*. 2013;208:1737–1746. doi: 10.1093/infdis/jit508
- 158. Kearns A, Gordon J, Burdo TH, Qin X. HIV-1–associated atherosclerosis: unraveling the missing link. *J Am Coll Cardiol*. 2017;69:3084–3098. doi: 10.1016/j.jacc.2017.05.012
- 159. Francisci D, Pirro M, Schiaroli E, Mannarino MR, Cipriani S, Bianconi V, Alunno A, Bagaglia F, Bistoni O, Falcinelli E, et al. Maraviroc intensification modulates atherosclerotic progression in HIV-suppressed patients at high cardiovascular risk. A randomized, crossover pilot study. *Open Forum Infect Dis*. 2019;6:ofz112. doi: 10.1093/ofid/ofz112
- 160. Cipriani S, Francisci D, Mencarelli A, Renga B, Schiaroli E, D'Amore C, Baldelli F, Fiorucci S. Efficacy of the CCR5 antagonist maraviroc in reducing early, ritonavir-induced atherogenesis and advanced plaque progression in mice. *Circulation*. 2013;127:2114–2124. doi: 10.1161/CIRCULATIONAHA.113.001278
- 161. Park IW, Wang JF, Groopman JE. HIV-1 Tat promotes monocyte chemoattractant protein-1 secretion followed by transmigration of monocytes. *Blood*. 2001;97:352–358. doi: 10.1182/blood.v97.2.352
- 162. Matzen K, Dirkx AEM, Oude Egbrink MGA, Speth C, Götte M, Ascherl G, Grimm T, Griffioen AW, Stürzl M. HIV-1 Tat increases the adhesion of monocytes and T-cells to the endothelium in vitro and in vivo: implications for AIDS-associated vasculopathy. *Virus Res*. 2004;104:145–155. doi: 10.1016/j.virusres.2004.04.001
- 163. Paladugu R, Fu W, Conklin BS, Lin PH, Lumsden AB, Yao Q, Chen C. Hiv Tat protein causes endothelial dysfunction in porcine coronary arteries. *J Vasc Surg*. 2003;38:549–55; discussion 555. doi: 10.1016/s0741-5214(03)00770-5
- 164. Meng Z, Hernandez R, Liu J, Gwag T, Lu W, Hsiai TK, Kaul M, Zhou T, Zhou C. HIV protein Tat induces macrophage dysfunction and atherosclerosis development in low-density lipoprotein receptor-deficient mice. *Cardiovasc Drugs Ther*. 2022;36:201–215. doi: 10.1007/s10557-021-07141-x
- 165. Swingler S, Mann A, Jacqué JM, Brichacek B, Sasseville VG, Williams K, Lackner AA, Janoff EN, Wang R, Fisher D, et al. HIV-1 Nef mediates lymphocyte chemotaxis and activation by infected macrophages. *Nat Med*. 1999;5:997–103. doi: 10.1038/12433
- 166. Cui HL, Ditiatkovski M, Kesani R, Bobryshev YV, Liu Y, Geyer M, Mukhamedova N, Bukrinsky M, Sviridov D. HIV protein Nef causes dyslipidemia and formation of foam cells in mouse models of atherosclerosis. *FASEB J*. 2014;28:2828–2839. doi: 10.1096/fj.13-246876
- 167. Wang S, Pasca S, Post WS, Langan S, Pallavajjala A, Haley L, Gocke CD, Budoff M, Haberlen S, Brown TT, et al. Clonal hematopoiesis in men living with HIV and association with subclinical atherosclerosis. *AIDS*. 2022;36:1521–1531. doi: 10.1097/QAD.0000000000003280
- 168. Alcaraz MJ, Alcaraz A, Teruel-Montoya R, Campillo JA, de la Torre A, Muñoz A, Tomás C, Puche G, Báguena C, Cano A, et al. Subclinical atherosclerosis and immune activation in young HIV-infected patients with telomere shortening. *Aging (Albany NY)*. 2021;13:18094–18105. doi: 10.18632/aging.203350
- 169. Bernal E, Martinez M, Torres A, Guillamón CF, Alcaraz A, Alcaraz MJ, Muñoz A, Valero S, Botella C, Campillo JA, et al. T cell senescence predicts subclinical atherosclerosis in HIV-infected patients similarly to traditional cardiovascular risk factors. *Antiviral Res*. 2019;162:163–170. doi: 10.1016/j.antiviral.2018.12.014
- 170. Palmer CS, Anzinger JJ, Zhou J, Gouillou M, Landay A, Jaworowski A, McCune JM, Crowe SM. Glucose transporter 1-expressing proinflammatory monocytes are elevated in combination antiretroviral therapy-treated and untreated HIV+ subjects. *J Immunol*. 2014;193:5595–5603. doi: 10.4049/jimmunol.1303092
- 171. Sviridov D, Miller YI, Bukrinsky MI. Trained immunity and HIV infection. *Front Immunol*. 2022;13:903884. doi: 10.3389/fimmu.2022.903884
- 172. Riksen NP, Bekkering S, Mulder WJM, Netea MG. Trained immunity in atherosclerotic cardiovascular disease. *Nat Rev Cardiol*. 2023;20:799–811. doi: 10.1038/s41569-023-00894-y
- 173. Funderburg NT, Jiang Y, Debanne SM, Labbato D, Juchnowski S, Ferrari B, Clagett B, Robinson J, Lederman MM, McComsey GA. Rosuvastatin reduces vascular inflammation and T-cell and monocyte activation in HIVinfected subjects on antiretroviral therapy. *J Acquir Immune Defic Syndr*. 2015;68:396–404. doi: 10.1097/QAI.0000000000000478
- 174. Friis-Møller N, Sabin CA, Weber R, d'Arminio Monforte A, El-Sadr WM, Reiss P, Thiébaut R, Morfeldt L, De Wit S, Pradier C, et al; Data Collection on Adverse Events of Anti-HIV Drugs (DAD) Study Group. Combination antiretroviral therapy and the risk of myocardial infarction. *N Engl J Med*. 2003;349:1993–2003. doi: 10.1056/NEJMoa030218
- 175. El-Sadr WM, Lundgren JD, Neaton JD, Gordin F, Abrams D, Arduino RC, Babiker A, Burman W, Clumeck N, Cohen CJ, et al; Strategies for Management of Antiretroviral Therapy (SMART) Study Group. CD4+ count-guided interruption of antiretroviral treatment. *N Engl J Med*. 2006;355:2283–2296. doi: 10.1056/NEJMoa062360
- 176. Lundgren JD, Babiker AG, Gordin F, Emery S, Grund B, Sharma S, Avihingsanon A, Cooper DA, Fätkenheuer G, Llibre JM, et al; INSIGHT START Study Group. Initiation of antiretroviral therapy in early asymptomatic HIV infection. *N Engl J Med*. 2015;373:795–807. doi: 10.1056/NEJMoa1506816
- 177. Friis-Møller N, Reiss P, Sabin CA, Weber R, Monforte A, El-Sadr W, Thiébaut R, De Wit S, Kirk O, Fontas E, et al; DAD Study Group. Class of

HIV AND CARDIOVASCULAR<br>Disease Review Series

antiretroviral drugs and the risk of myocardial infarction. *N Engl J Med*. 2007;356:1723–1735. doi: 10.1056/NEJMoa062744

- 178. Bavinger C, Bendavid E, Niehaus K, Olshen RA, Olkin I, Sundaram V, Wein N, Holodniy M, Hou N, Owens DK, et al. Risk of cardiovascular disease from antiretroviral therapy for HIV: a systematic review. *PLoS One*. 2013;8:e59551. doi: 10.1371/journal.pone.0059551
- 179. Coplan PM, Nikas A, Japour A, Cormier K, Maradit-Kremers H, Lewis R, Xu Y, DiNubile MJ. Incidence of myocardial infarction in randomized clinical trials of protease inhibitor-based antiretroviral therapy: an analysis of four different protease inhibitors. *AIDS Res Hum Retroviruses*. 2003;19:449– 455. doi: 10.1089/088922203766774487
- 180. Ryom L, Lundgren JD, El-Sadr W, Reiss P, Kirk O, Law M, Phillips A, Weber R, Fontas E, d' Arminio Monforte A, et al; D:A:D study group. Cardiovascular disease and use of contemporary protease inhibitors: the D:A:D international prospective multicohort study. *Lancet HIV*. 2018;5:e291–e300. doi: 10.1016/S2352-3018(18)30043-2
- 181. Monforte A, Reiss P, Ryom L, El-Sadr W, Dabis F, De Wit S, Worm SW, Law MG, Weber R, Kirk O, et al. Atazanavir is not associated with an increased risk of cardio- or cerebrovascular disease events. *AIDS*. 2013;27:407–415. doi: 10.1097/QAD.0b013e32835b2ef1
- 182. Worm SW, Sabin C, Weber R, Reiss P, El-Sadr W, Dabis F, De Wit S, Law M, Monforte AD, Friis-Møller N, et al. Risk of myocardial infarction in patients with HIV infection exposed to specific individual antiretroviral drugs from the 3 major drug classes: the data collection on adverse events of anti-HIV drugs (D:A:D) study. *J Infect Dis*. 2010;201:318–330. doi: 10.1086/649897
- 183. O'Halloran JA, Dunne E, Tinago W, Denieffe S, Kenny D, Mallon PWG. Switching from abacavir to tenofovir disoproxil fumarate is associated with rises in soluble glycoprotein VI, suggesting changes in platelet–collagen interactions. *AIDS*. 2018;32:861–866. doi: 10.1097/QAD.0000000000001783
- 184. Baum PD, Sullam PM, Stoddart CA, McCune JM. Abacavir increases platelet reactivity via competitive inhibition of soluble guanylyl cyclase. *AIDS*. 2011;25:2243–2248. doi: 10.1097/QAD.0b013e32834d3cc3
- 185. Hsue PY, Hunt PW, Wu Y, Schnell A, Ho JE, Hatano H, Xie Y, Martin JN, Ganz P, Deeks SG. Association of abacavir and impaired endothelial function in treated and suppressed HIV-infected patients. *AIDS*. 2009;23:2021–2027. doi: 10.1097/QAD.0b013e32832e7140
- 186. Ding X, Andraca-Carrera E, Cooper C, Miele P, Kornegay C, Soukup M, Marcus KA. No association of abacavir use with myocardial infarction: findings of an FDA meta-analysis. *J Acquir Immune Defic Syndr*. 2012;61:441– 447. doi: 10.1097/QAI.0b013e31826f993c
- 187. Vos AG, Venter WDF. Cardiovascular toxicity of contemporary antiretroviral therapy. *Curr Opin HIV AIDS*. 2021;16:286–291. doi: 10.1097/COH.0000000000000702
- 188. Neesgaard B, Greenberg L, Miró JM, Grabmeier-Pfistershammer K, Wandeler G, Smith C, De Wit S, Wit F, Pelchen-Matthews A, Mussini C, et al. Associations between integrase strand-transfer inhibitors and cardiovascular disease in people living with HIV: a multicentre prospective study from the RESPOND cohort consortium. *Lancet HIV*. 2022;9:e474–e485. doi: 10.1016/S2352-3018(22)00094-7
- 189. Surial B, Chammartin F, Damas J, Calmy A, Haerry D, Stöckle M, Schmid P, Bernasconi E, Fux CA, Tarr PE, et al; Swiss HIV Cohort Study. Impact of integrase inhibitors on cardiovascular disease events in people with human immunodeficiency virus starting antiretroviral therapy. *Clin Infect Dis*. 2023;77:729–737. doi: 10.1093/cid/ciad286
- 190. Piñeirua-Menéndez A, Flores-Miranda R, Sánchez-Nava D, Ortega-Pérez R, Belaunzaran-Zamudio PF, Pérez-Patrigeon S, Cárdenas-Ochoa A, Oseguera-Moguel J, Galindo-Uribe J, Orihuela-Sandoval C, et al. Myocardial inflammatory changes before and after antiretroviral therapy initiation in people with advanced HIV disease. *Open Forum Infect Dis*. 2020;7:ofaa297. doi: 10.1093/ofid/ofaa297
- 191. Schuster C, Mayer FJ, Wohlfahrt C, Marculescu R, Skoll M, Strassl R, Pavo N, Popow-Kraupp T, Hülsmann M, Bauer M, et al. Acute HIV infection results in subclinical inflammatory cardiomyopathy. *J Infect Dis*. 2018;218:466–470. doi: 10.1093/infdis/jiy183
- 192. Zanni MV, Toribio M, Robbins GK, Burdo TH, Lu MT, Ishai AE, Feldpausch MN, Martin A, Melbourne K, Triant VA, et al. Effects of antiretroviral therapy on immune function and arterial inflammation in treatment-naive patients with human immunodeficiency virus infection. *JAMA Cardiol*. 2016;1:474–480. doi: 10.1001/jamacardio.2016.0846
- 193. McLaughlin MM, Ma Y, Scherzer R, Rahalkar S, Martin JN, Mills C, Milush J, Deeks SG, Hsue PY. Association of viral persistence and atherosclerosis in adults with treated HIV infection. *JAMA Netw Open*. 2020;3:e2018099. doi: 10.1001/jamanetworkopen.2020.18099
- 194. Baker JV, Henry WK, Patel P, Bush TJ, Conley LJ, Mack WJ, Overton ET, Budoff M, Hammer J, Carpenter CC, et al; Study to Understand the Natural History of HIV/AIDS in the Era of Effective Therapy Investigators. Progression of carotid intima-media thickness in a contemporary human immunodeficiency virus cohort. *Clin Infect Dis*. 2011;53:826–835. doi: 10.1093/cid/cir497
- 195. Post WS, Haberlen S, Witt MD, Zhang L, Jacobson LP, Brown TT, Margolick JB, Kingsley L, Palella FJ Jr, Budoff M. Suboptimal HIV suppression is associated with progression of coronary artery stenosis: the Multicenter AIDS Cohort Study (MACS) Longitudinal Coronary CT Angiography Study. *Atherosclerosis*. 2022;353:33–40. doi: 10.1016/j.atherosclerosis.2022.04.019
- 196. Cohn LB, Chomont N, Deeks SG. The biology of the HIV-1 latent reservoir and implications for cure strategies. *Cell Host Microbe*. 2020;27:519–530. doi: 10.1016/j.chom.2020.03.014